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Citation: Cima Cabal MD, Molina F, López-Sánchez JI, Pérez-Santín E, del Mar García-Suárez M (2023) Pneumolysin as a target for new therapies against pneumococcal infections: A systematic review. PLoS ONE 18(3): e0282970. https://doi.org/ 10.1371/journal.pone.0282970

Editor: Raj Kumar Koiri, Dr. Harsingh Gour Central University, INDIA

Received: October 19, 2022

Accepted: February 28, 2023

Published: March 22, 2023

Peer Review History: PLOS recognizes the benefits of transparency in the peer review process; therefore, we enable the publication of all of the content of peer review and author responses alongside final, published articles. The editorial history of this article is available here: https://doi.org/10.1371/journal.pone.0282970

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Data Availability Statement: All relevant data are within the paper and its <u>Supporting Information</u> files.

RESEARCH ARTICLE

Pneumolysin as a target for new therapies against pneumococcal infections: A systematic review

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Abstract

Background

This systematic review evaluates pneumolysin (PLY) as a target for new treatments against pneumococcal infections. Pneumolysin is one of the main virulence factors produced by all types of pneumococci. This toxin (53 kDa) is a highly conserved protein that binds to cholesterol in eukaryotic cells, forming pores that lead to cell destruction.

Methods

The databases consulted were MEDLINE, Web of Science, and Scopus. Articles were independently screened by title, abstract, and full text by two researchers, and using consensus to resolve any disagreements that occurred. Articles in other languages different from English, patents, cases report, notes, chapter books and reviews were excluded. Searches were restricted to the years 2000 to 2021. Methodological quality was evaluated using OHAT framework.

Results

Forty-one articles describing the effects of different molecules that inhibit PLY were reviewed. Briefly, the inhibitory molecules found were classified into three main groups: those exerting a direct effect by binding and/or blocking PLY, those acting indirectly by preventing its effects on host cells, and those whose mechanisms are unknown. Although many molecules are proposed as toxin blockers, only some of them, such as antibiotics, peptides, sterols, and statins, have the probability of being implemented as clinical treatment. In contrast, for other molecules, there are limited studies that demonstrate efficacy in animal models with sufficient reliability.

Discussion

Most of the studies reviewed has a good level of confidence. However, one of the limitations of this systematic review is the lack of homogeneity of the studies, what prevented to carry out a statistical comparison of the results or meta-analysis.

Funding: This work was supported by Universidad Internacional de La Rioja under the project Pneumo-SARS UNIR-B0036 (2021–2022). The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Competing interests: The authors declare no competing interest.

Conclusion

A panel of molecules blocking PLY activity are associated with the improvement of the inflammatory process triggered by the pneumococcal infection. Some molecules have already been used in humans for other purposes, so they could be safe for use in patients with pneumococcal infections. These patients might benefit from a second line treatment during the initial stages of the infection preventing acute respiratory distress syndrome and invasive pneumococcal diseases. Additional research using the presented set of compounds might further improve the clinical management of these patients.

Introduction

Streptococcus pneumoniae is the leading cause of community-acquired pneumonia in both adults and children [1]. It is also a common cause of meningitis and septicemia as well as other minor infections such as sinusitis and otitis media [2], even despite the extensive vaccination programs that exist today, especially in developed countries [3]. The prevalence of this bacterium is associated with its virulence factors and the patient's own risk factors including age, smoking, and other types of comorbidities (*e.g.*, diabetes or immunodeficiency). Virulence of *S. pneumoniae* is due to several factors, some of them related to the structure of the bacteria, such as the capsular polysaccharide (whose differences are related to bacteria serotypes) and the variety of surface protein families (*i.e.*, lipoproteins, sortase-anchored proteins, choline-binding proteins, and the non-classical surface proteins), but also the cytoplasmatic toxin, pneumolysin (PLY). The nature and action modes of these virulence factors, directly involved in the pathogenicity of pneumococcus, have been previously described elsewhere [1, 2, 4–6].

PLY is a toxin that binds to eukaryotic membrane cholesterol (belongs to cholesteroldependent cytolysins, CDC) but also binds to the mannose receptor C type 1 (MRC-1) promoting an anti-inflammatory response and reducing pneumococcal disease [7, 8]. In this way, this toxin has double functionality ("sword and shield" or "Yin and Yang") [9]. PLY is produced constitutively but free toxin is higher in the late log phase due to the presence of a defined threshold concentration of extracellular autolysin (LytA) which dictates the onset of autolysis. The entry into the stationary phase due to nutrient depletion sensitizes cells to the effect of LytA, while during exponential growth they are protected from the action of this enzyme [10, 11]. Other investigations have also revealed the release of PLY in the extracellular vesicles [12]. On the other hand, a phenotypic heterogenicity has been demonstrated in terms of the level of expression of PLY, which helps the dispersion of the pneumococcus through the host [9, 13]. PLY does not have attachment motifs, however the toxin localizes to the cell envelope of actively growing cells, where its release and activity is controlled by the composition of the peptidoglycan, specifically by the proportion of branched stem peptides that vary throughout the cell cycle and between different strains [14].

PLY belongs to the family of thiol-activated toxins, commonly produced by many Grampositive bacteria, which creates membrane pores in eukaryotic cells by binding to cholesterol, thus causing cell destruction [15]. It is worth noting that serotypes 1 and 8 have been shown to harbor mutations in the *ply* gene that annul this main characteristic and cause a much milder disease, due to a non-hemolytic allele (sequence type, ST306) allows adaptation to an intracellular lifestyle [16, 17]. However, in sub-Saharan Africa, serotype 1 causes invasive pneumococcal disease due to an increased production of autolysin and hemolytic pneumolysin alleles [18]. (ST217). This serotype is a major cause of invasive pneumococcal disease globally, especially in Africa, South America, and Asia, with geographically distinct sequence types (STs) that form three genetic clusters designated as lineage A, B, and C [19].

Structurally, PLY consists of 471 amino acids (53 kDa) and has a three-dimensional conformation with 4 different domains; three of them (domains 1, 2, and 3) have structural importance, conferring stability to the PLY molecule, and are essential for oligomerization and pore formation in eukaryotic cell membranes. Domain 4 (hydrophobic region, loops L1-L3) forms the C-terminal region and is what promotes cholesterol binding, favoring the insertion of an undecapeptide sequence in the membrane, which allows for the oligomerization of toxin monomers, and the further formation of pores. The PLY pore is a 400 Å ring of 42 membraneinserted monomers [20]. (S1a Fig) [21, 22].

This toxin can activate different cell death pathways such as apoptosis [23], pyroptosis [24], or necroptosis [25], which releases membrane-derived vesicles (microvesicles and exosomes). The pores of PLY cause a strong mitochondrial calcium influx which triggers mitochondrial morphological alterations with the release of mtDNA through microvesicles [26] and could regulate innate immune responses [27] (S1a Fig). During the inflammatory process, PLY activates several signal transduction pathways such as the nuclear factor kappa-B (NF- κ B), mitogen-activated protein kinase (MAPK), and the NOD-like receptor pyrin domain-containing 3 (NLRP3) inflammsone [28, 29]. The interaction between PLY and TLR4 remains controversial. Some authors showed that the interaction of PLY with TLR4 is involved in induce cytokine production and apoptosis [30, 31], while other research showed that PLY activates the NLRP3/ACS inflammasome to enhance the secretion of pro-inflammatory cytokines IL-1 β and IL-18 from macrophages and dendritic cells and contributes to the protection of the host from pneumococcal infection independent of TLR-4 and mediated by K⁺ influx [24, 32].

Therefore, PLY is able to activate and regulate a huge number of genes for chemokines, cytokines, and other molecules whose expression is involved in the recruitment of inflammatory cells via neutrophils, macrophages, and phagocytes activation (such as IL-8, MCP-3, MIP-1 β , lysozymes, caspases, TNF, IL-1 and IL-6) [33, 34]. Overproduction of early cytokines has been associated with tissue injury, organ dysfunction, morbidity, and mortality [35].

On the other hand, PLY is the main cause of pulmonary permeability edema due to its ability to alter both endothelial and epithelial barrier function [36]. This effect is a consequence of reducing dynamic and stable microtubule content in the endothelial monolayer and influencing VE-cadherin expression [37]. NLRP3 protect the alveolar barrier againts PLY injury [38]. Several cellular channels or transporters of the alveolar epithelium are implicated, such as the epithelial sodium channel (ENaC), the Na⁺/ K⁺ -ATPase and K⁺ channels [39] (S1a Fig).

Moreover, PLY play a key role in human nasopharynx colonization and in the transmission of *S. pneumoniae* from host to host since by promoting inflammation there is an increase in elimination [40]. In addition, *S. pneumoniae* can invade other parts of the body via blood-stream dissemination, and it can gain access to normally 'sterile' sites such as the lower airways or meninges [41]. (S1b Fig). When bacteria reach the cerebrospinal fluid, produce meninges inflammation, resulting in hyperemia and ischemia, and eventually permanent brain injury [42].

Finally, it has been found that CAP (community acquired pneumonia) can incite up to 30% of the cases to cardiovascular events, such as myocardial damage or pro-thrombotic effects [43]. Myocardial damage is closely related to the ability of PLY to form pores in the cell membrane: movement of Ca²⁺ into the cell leads to an efflux of K⁺ with the consequent depolarization of the membrane, contributing to myocardial contractile dysfunction [44]. Regarding prothrombotic effects, this has recently been shown not to be true. PLY does not activate

platelets to form thrombus, rather it destroys them by forming pores in their membrane [45] and destroy procoagulant microvesicles impaired coagulation of blood [46].

Current pneumococcal conjugate vaccines are 13-valent conjugate vaccine (PCV13) for routine pediatric immunization and a 23-valent polysaccharide vaccine (PPSV23) for adults aged \geq 65 years. Since 2014, PCV13 was also recommended for all adults aged \geq 65 years [47]. In 2021, two new vaccines were approved by the FDA for use in adults \geq 18 years (PCV-15 and PCV-20) that are presently under evaluation [48]. Although these vaccines are immunogenic and effective and prevent disease caused by the serotypes determined by their capsule types, they do not cover the full spectrum of invasive pneumococcal serotypes. The management of pneumococcal infections in clinical practice frequently involves the use of broad-spectrum antibiotics, typically a combined therapy of β -lactams and macrolides [49]. However, *S. pneumoniae* has developed resistance to multiple antibiotics including penicillin, macrolides, fluoroquinolone, and sulfamethoxazole-trimethoprim [50]. The emergence of non-vaccine serotypes after the introduction of PCV, together with increased antibiotic resistance in these serotypes, has become a global threat [51]. Taking together the development of new therapies is necessary for the effective treatment of pneumococcal infections.

Even though PLY-neutralization strategies have been reviewed before, these are partial reviews, that approach the subject from different perspectives, and the vast majority of these reviews focus on vaccination-based strategies (see for example [3, 52–56]). Furthermore, several previous reviews have been carried out on small-molecule-based PLY neutralization strategies that either suffer from being focused on a single family of compounds and do not represent complete overviews of the current state of the art, such as the work of Nishimoto et al. (2020) [57] which is focused on statins, the work of Li et al. (2017) [58], focused on sterrols, or the work of Anderson and Feldman (2017) [59], which although it encompasses a broader collection of compounds, it cannot be considered complete and current to date.

The aim of this review was to investigate the main molecules that directly and indirectly interfere with PLY activity in host survival, colonization, infection, and transmission. A secondary objective was to identify the mechanisms of interaction with the toxin.

Materials and methods

This systematic review was conducted in accordance with the Preferred Reporting Items for Systematic Reviews (PRISMA) guidelines [60] as much as possible, since it is not a medical systematic review, so part of the guidelines were not applicable (S1 Table).

Search strategy and selection criteria

The inclusion criteria of the studies for the review were articles in which PLY was examined as a therapeutic target through studies of molecules that inhibit its effects. To analyze the therapeutic role of PLY in pneumococcal infection, the terms "pneumolysin" AND "therapeutic" were searched in three databases: Web of Science, MEDLINE and Scopus. At first, we only limited the search date to November 25, 2021; however, in the exclusion criteria, we decided to refine and not include articles prior to the year 2000. Therefore, the review includes the articles found from January 1st, 2000 to November 25th, 2021. Reference lists of selected articles were also reviewed. Several exclusion criteria were taken to ensure the quality of the study. The following were excluded: patents, articles in other languages different from English, cases report, notes, chapter books, reviews, and articles prior to 2000 year. Potentially eligible articles were reviewed by two independent reviewers (MMG-S and MDCC). Disagreements between the two reviewers were resolved by consensus.



PRISMA 2020 flow diagram for new systematic reviews which included searches of databases, registers and other sources

*Consider, if feasible to do so, reporting the number of records identified from each database or register searched (rather than the total number across all databases/registers) **If automation tools were used, indicate how many records were excluded by a human and how many were excluded by automation tools.

From: Page MJ, McKenzie JE, Bossuyt PM, Boutron I, Hoffmann TC, Mulrow CD, et al. The PRISMA 2020 statement: an updated guideline for reporting systematic reviews. BMJ 2021;372:n71. doi: 10.1136/bmj.n71. For more information, visit: http://www.prisma-statement.org/

Fig 1. Flowchart of the systematic review process.

https://doi.org/10.1371/journal.pone.0282970.g001

Selection procedure

Citations were saved in the reference manager EndNote (https://endnote.com/) and in an Excel file that also included the following items: publication type, authors, article title, abstract, source title, DOI, and publication date. EndNote was used to deduplicate the search results. Full texts of these articles were analyzed by the two authors (MMG-S and MDCC). Articles on the pathogenesis of pneumococcus in general and articles in which PLY was presented as a vaccine and as tumor therapy were excluded from the study. Fig 1 shows an overview of process used for the selection of the articles included in this review.

Data collection process

Two authors collected data from articles in an electronic spreadsheet (**S2 Table**). The following data were obtained from each article: tests performed on cell lines and laboratory animals, CAS number and tested doses of the compound, route of inoculation, type of infection, and strain of *S. pneumoniae* used as well as other data considered of interest.

Data analysis and synthesis

Quantitative and qualitative outcome data were provided and synthesized where possible. Due to data heterogeneity a meta-analysis was not possible.





https://doi.org/10.1371/journal.pone.0282970.g002

Article quality assessment

Each article was scrutinized to determine the techniques that led to each molecule being proposed as a candidate for toxin-blocking therapy. The risk assessment tool used was the Office of Health Assessment & Translation (OHAT) risk of bias tool by the US Department of Health and Human Services [61]. Risk assessment was based on study type. This screening was tabulated in <u>S3 Table</u>. Moreover, we also inspected the number of papers of clinical trials indexed in MEDLINE for each proposed molecule (<u>S4 Table</u>; Fig 2).

Network of neutralizing PLY-effects molecules and mechanisms underlying

The knowledge network harboring the molecules, the processes affected, and the cellular targets studied in this review were all generated using the tool EXTRACT 2.0 [62] to gather the terms from the articles' abstracts. From the resulting dataset, a graph was built using Cytoscape 3.9.1 [63], using the yFiles layout algorithms. Finally, some images were obtained from BioRender.com (2022).

Results and discussion

Study selection

A total of 366 records were found using the search criteria, of which 203 were found in WOS, 161 in MEDLINE and 2 in Scopus (Fig 1). Once the duplicates were eliminated, 192 articles

were reviewed by title and abstract by two independent authors (MG and MC). Several exclusion criteria were taken into account to ensure the review quality. As a result, 25 reviews, 16 articles, 3 patents, 2 cases report, 2 articles in other languages different than English, 2 chapter books, and 1 note prior to the year 2000 were excluded. The full texts of the resulting 141 articles were reviewed in detail. This refined search showed that 108 studies did not meet the criteria and were excluded (74 reports about the effect of PLY as a vaccine, 3 reports focused on tumor therapy, 31 reports studying general pathogenesis). References from these relevant articles were also screened. Forty-one (n = 41) studies met the inclusion criteria (describing molecules that inhibited the effects of PLY) were included in the systematic review.

Study characteristics

A detailed analysis of the 41 resulting articles was performed, in which the effects of PLY were studied using molecules that block it. Most of the studies were published by scientists from Southeast Asia, principally China (n = 17), followed by Europe (n = 10), and the USA (n = 9) (**S4 Table**). In 15 of the articles, the experiments were carried out using only cell cultures, while in 26, animal models of pneumococcal infection were used (24 articles induced a pneumonia model, of which two also induced systemic infection) [64, 65], one model of meningitis [66], two of keratitis [67, 68], one of atherosclerosis [69] and one of nasopharyngeal colonization [70]. The most frequently used animal models were mice for pneumonia and rabbits for keratitis. Meningitis was tested in rats and systemic infections were tested in Zebrafish embryos.

In 17 articles, the cellular cultures used were the human lung alveolar epithelial cell line A549, the human bronchial epithelial cell line HBE1 and H441 (n = 2), mouse leukemic monocyte-macrophage cell line RAW264.7 (n = 3), human umbilical vein endothelial cells (n = 3), human lung microvascular endothelial cells (n = 2), neutrophils (n = 2), cochlear hair cells (n = 1), primary glial cultures (n = 1), and primary astrocytes (n = 1). Laboratory animals used to induce pneumococcal pneumonia were C57BL/6 mice (n = 12), BALB/c mice (n = 8), MF-1 mice (n = 1), ICR mice (n = 1), and C3H mice (n = 1).

Methodological quality

The OHAT quality scores of the included articles are described in **S3 Table**. The overall risk of bias rating was considered as Low or Probably Low. The 41 studies were summarized and classified into 10 groups according to chemical structures: Plant-derived compounds, sterols, statins, omega-3 fatty acids, purin-6-ones, thioethers, antibiotics, peptides, cations, and antibodies (**Table 1**). Moreover, because of the high heterogeneity of the data found in the articles, it was very difficult to compare the action mechanism against the toxin from each study, so they were grouped into three categories for clarification: direct (the molecule binds to or competes with PLY for binding to the target), indirect (the molecule does not bind PLY), and unknown (molecular mechanisms are currently unknown).

On the other hand, Fig 2 shows the number of articles indexed as 'clinical trials' on MED-LINE (without time restrictions), illustrating that only 17 of 39 molecules present experimental human evidence to potentially conclude data on their efficacy, toxicity, or bioavailability. The distribution of studies regarding molecules is unequal. For some molecules, there are more than 1000 studies (like cases of calcium and cholesterol), for others, the number ranges from 50–100, and for 10 of them, there are less than 20 studies. As expected, more trials and global studies are found to specifically study sepsis and pneumonia molecules. Thus, most trials focused on sepsis are limited to molecules, exhibiting more than 1000 global studies.

Reference	Molecule (No CAS)	Molecule type	Toxicity assays	Action mechanism	Infection type	Cellular lines and	S. pneumoniae
[77]	Acacetin (480-44-4)	Flavonoid, polyphenol (natural)	Cytotoxicity assays: Purified PLY (0.4 μ M) was incubated with different concentrations of acacetin, 8, 16, 32 μ g/ml, all of them significantly decreased the LDH release of A549 cells. Animal models: Intranasal infected mice were injected subcutaneously with acacetin at 50 mg/kg. The levels of IL-1 β and IFN- γ in bronchoalveolar lavage fluid and the wet/dry weight ratio of lung tissue was significantly decreased at 96 h post- infection.	Direct . Acacetin inhibit the oligomerization of PLY. Iteration residues no shown.	Pneumonia	laboratory animals BALB/c mice	strain D39
[78]	Amentoflavone (1617-53-4)	Flavonoid, polyphenol (natural)	Cytotoxicity assays: A549 cells were treated with PLY (0.825 µg/ml) and increasing concentrations of amentoflavone (AMF). LDH release significantly decreased at AMF concentrations higher than 2 µg/ ml. Animal models: Mice were administered with AMF (50 mg/kg) subcutaneously 2 h after infection with D39. Analysis of lungs 72 h after infection showed significant improvement of inflammatory cell accumulations and alveolar damage with respect to the control mice.	Direct . Amentoflavone interacts with the toxin at Ser254, Glu277, Arg359, and weakens the oligomerization of PLY.	Pneumonia	A549 cells ¹ C57BL/ 6J mice	D39
[79]	Morin (480-16-0)	Flavonoid, polyphenol (natural)	Cytotoxicity assays: Morin reduces the cytotoxicity of PLY (0.2 mg/ml) in cells at concentrations above 2 µg/ml. Animal models: Infected mice were injected subcutaneously with 50 mg/kg morin two hours after infection. Three days post-infection, the lung tissues in the morin treatment group of mice displayed a significant reduction of inflammatory cell infiltration, bacteria number, and pulmonary inflammation.	Direct. Morin inhibits the oligomerization of PLY. Iteration residues no shown.	Pneumonia	A549 cells C57BL/6J mice	D39
[76]	Apigenin (520-36-5)	Flavonoid, polyphenol (natural)	Cytotoxicity assays: A549 cells were exposed to 80 nM PLY and different concentrations of apigenin (0, 10, 20, 40 and 80 μ M). Apigenin at a concentration range of 40 to 80 μ M conferred significant protection against toxin cellular effects. Animal models: Mice were intranasally infected and 2 h later subcutaneously administered apigenin (80 mg/kg). At 48 h post-infection, the bacterial burden, the tumor necrosis factor α (TNF- α) and interleukin 1 β (IL-1 β) in the bronchoalveolar lavage fluid levels were significantly reduced compared with the levels detected in the control mice.	Direct. Apigenin Inhibits the oligomerization of PLY. Iteration residues no shown.	Pneumonia	A549 cells BALB/c mice	D39
[80]	Epigallocatechin gallate (989-51-5)	Flavonoid, Polyphenol (natural)	Cytotoxicity assays: A549 cells were pre-incubated with various concentrations of Epigallocatechin gallate (EGCG) (0, 1.09, 2.18, 4.36, and 8.73 µM) and incubated with PLY (80 nM). Concentrations above 1.09 µM showed significant differences in LDH release. Animal models: Mice were intranasally inoculated to produce pneumonia and were subcutaneously administered EGCG (50 mg/kg). The mortality at 120 h decreased approximately a 40%.	Direct . Epigallocatechin gallate inhibits the oligomerization of PLY. The binding model of EGCG with PLY revealed that the side chain of EGCG can form strong interactions with Ser256, Glu277, Tyr358 and Arg359.	Pneumonia	A549 cells Hep2 ² BALB/c mice	D39
[81]	Quercetin (117-39-5)	Flavonoid	Cytotoxicity assays: A549 cells incubated with 3 µl PLY (0.16 mg/ml) and preincubated with Quercetin (2, 4, 8, 16, 32 µg/ml) significantly reduced PLY cytotoxicity at all concentrations assayed. Animal models: Infected mice were subcutaneously treated with Quercetin (25 mg/kg). After 96h post- infection, the survival rate of treated mice was 80% versus 60% in control mice. Edema and cytokine release were decreased.	Direct. Quercetin inhibits PLY oligomerization. Iteration residues no shown.	Pneumonia	A549 cells BALB/c	D39
[82]	Dryocrassin ABBA (12777-70-7)	Flavonoid	Hemolysis: PLY (0.4 μM) was incubated with ABBA (0, 1, 2, 4, and 8 μg/ml). Reduction of hemolytic activity was observed at concentrations above 2 μg/ml. Cytotoxicity assays: Cells incubated with ABBA (2, 4, 8, 16 μg/ml) and PLY (0.4 μM) reduced PLY toxicity at 16 μg/ml.	Direct. Quercetin inhibits PLY oligomerization. Iteration residues no shown.		RAW264.7 cells ³	D39
[75]	Verbascoside (61276-17-3)	Phenylpropanoid glycoside, Polyphenol (natural)	Cytotoxicity assays: LDH release assay was performed to evaluate the effect of Verbascoside (VBS) on PLY-mediated lysis of A549 cells. The addition of 2 to 32 µg/ml of VB) reduced the cytotoxicity of PLY in a dose-dependent manner. Animal models: Mice infected with <i>S.</i> <i>pneumoniae</i> were treated with 100 mg/kg VBS. Survival rate at 120 h after infection was 75% comparing with the 20% untreated control group.	Direct. Verbascoside inhibits the oligomerization of PLY binds to the cleft between domains 3 and 4 of PLY. The hydroxyl group of the benzene ring on the right side of VBS can form a strong hydrogen bond with Asp471 which has a crucial significance in making the right side of VBS stable.	Pneumonia	A549 cells C57BL/6J mice	D39

Table 1. Basic characteristics of the articles included into this report show the molecules with effects against PLY.

Reference	Molecule (No CAS)	Molecule type	Toxicity assays	Action mechanism	Infection type	Cellular lines and laboratory animals	S. pneumoniae strain
[83]	Pentagalloylglucose (14937-32-7) and gemin A (82220-61-9) $\underline{C}_{82}\underline{H}_{56}\underline{O}_{52}$	Tannins, polyphenols (natural)	$\label{eq:hardward} \begin{array}{l} \mbox{Hemolysis: Inhibitory activity of PLY was} \\ \mbox{calculated. The Pentagalloylglucose (PGG) IC_{50} was} \\ \mbox{18 \pm 0.7 nM and gemin A IC_{50} was 41 \pm 1 nM.} \\ \mbox{Cytotoxicity assays: A549 cells were incubated with} \\ \mbox{2 nM of PLY and 500, 1000 and 2000 nM of PGG} \\ \mbox{inhibited LDH release by 60\%, 87\% and 90\%,} \\ \mbox{respectively.} \end{array}$	Direct. 27 hydrolysable tannins were tested. PGG and gemin A were the most active monomer and oligomer, respectively. PLY oligomerization on the erythrocyte surface was inhibited with PGG. Flexible galloyl groups of PGG binds to the pocket formed by domains 2, 3, and 4 of PLY (Glu42, Ser256, Asp257, Glu277 and Arg359).		A549 cells	
[85]	Juglone (481-39-0)	Naphthoquinone	Hemolysis: 4nM PLY was mixed with Juglone (JG) (0, 2.6, 5.2, 10.4 and 20.8 µg/ml). Hemolytic activity was significantly reduced at 10.4 µg/ml. Cytotxicity assays: Cells were incubated with 80 nM PLY and JG (0, 2.6, 5.2, 10.4 and 20.8 µg/ml). Significant reduction of cytotoxicity was observed at 10.4 µg/ml.	Direct. Juglone inhibits the oligomerization of PLY. Iteration residues no shown.		A549 cells	D39
[84]	Shikonin (517-89-5)	Naphthoquinone (natural)	Cytotoxicity assays: The A549 cells were combined with different concentrations of shikonin (0.5– 16 µg/ml and PLY (0.2 µg). All concentrations of shikonin inhibited the toxic effect of PLY. Animal models: Micc with endonasal pulmonary infection were orally treated with shikonin (50 mg/ kg). 5 days after infection, the percentage of survival in treated mice was 65% versus 10% in the control mice.	Direct. Shikonin inhibits the oligomerization of PLY. Shikonin could bind to PLY through contact with certain amino acid sites thus affecting the conformational transition for PLY from the monomeric to oligomeric form.	Pneumonia	A549 cells C57BL/6J mice	D39
[71]	Aloe-emodin (481-72-1)	Anthraquinone (natural)	Hemolysis: S. pneumoniae cultures were treated with light alone (72 J/cm2) (P-L+), Aloe-emodin (AE) (32 µg/ml) alone (P+L-) or AE combined with light (P+L+) and supernatants added to erythrocytes. The hemolytic activity diminished 93.67% (P-L-) 83.33% (P-L+) 63.67% (P+L-) and 23.66% (P+L+).	Unknown. AE+ photodynamic therapy (435 ± 10 nm) reduced bacterial survival, biofilm formation, cytokine production, and inhibits PLY expression.		RAW264.7 cells	ATCC 49619 MDR (ATCC 49619
[86]	Ephedrine hydrochloride (50-98-6) Pseudoephedrine hydrochloride (345-78-8)	Phenethylamines (natural)	Cytotoxicity assays : A549 cells were cocultured with Ephedrine hydrochloride ESG (0, 4, 8, 16 and 32 µg/ml) preincubated with PLY (0.2 µg). LDH released was significantly reduced at 4 µg/ml. Animal models : In vivo protection was tested in infected mice at 72h. In mice subcutaneously treated with ESG (40 mg/kg), the percentage of survival was 50% versus 0% in the untreated control group.	Direct. ESG inhibits the oligomerization of PLY. Iteration residues no shown.	Pneumonia	A549 cells BALB/c mice	D39
[87]	Ephedrine hydrochloride (50-98-6) Pseudoephedrine hydrochloride (345-78-8) Methylephedrine (552-79-4) Amygdalin (2983-15-6) Prunasin (99-18-3) Glycyrrhetinic acid (471-53-4)	Phenethylamines (natural) Amygdalin and Glycyrrhetinic acid cyanogenic glycoside Glycyrrhetinic acid steroid hormone	Cytotoxicity assays: A549 cells were pre-incubated with various concentrations of MXSGT (0, 4, 8, 16 or 32 µg/ml) and incubated with PLY (0.2 µg/ml). Concentrations above 8 µg/ml showed significant differences in LDH release compared with the control. Animal models: Percentage of survival of mice infected and treated with MXSGT (50 mg/kg) was 40% protection at 72h versus 0% in the untreated mice control group.	Direct. Inhibits the oligomerization of PLY. Iteration residues no shown.	Pneumonia	A549 cells BALB/c mice	D39
[91]	Hederagenin (465-99-6)	Triterpenoid	Hemolysis: Hederagenin (2–32 μg/ml) was incubated with PLY (0.037 mg/ml). Hederagenin significantly inhibited the hemolytic activity of PLY at 8 μg/ml. Cytotoxicity assays: The protector effect was observed at concentrations above 16 μg/ml in cells incubated with hederagenin (0, 2, 8 and 32 μg/ml) and PLY.	Direct. Hederagenin interfered with the oligomerization of PLY. Iteration residues no shown.		A549 cells RAW264.7 cells	D39

Reference	Molecule (No CAS)	Molecule type	Toxicity assays	Action mechanism	Infection type	Cellular lines and laboratory animals	S. pneumoniae strain
[90]	Betulin (473-98-3)	Triterpene	Hemolysis: 2 µl of PLY (0.1 mg/ml) was incubated with Betulin (0,1, 2, 4, 8 µg/ml). Inhibition of hemolysis was observed at concentrations above 2 µg/ml. Cytotoxicity assays: A549 cells were exposed to PLY (80 nmol/l) and Betulin (4, 8, 16, 32 µg/ml). Protection of cytotoxicity was dose dependent. DNA-damage was also protected above 8 µg/ml.	Direct . Betulin inhibited the oligomerization of PLY. Iteration residues no shown.		A549 cells	D39
[89]	Oleanolic Acid (508-02-1)	Pentacyclic triterpenoids (natural)	Hemolysis: PLY was mixed with different concentrations of Oleanolic Acid (OA) (from 0 to 32 µg/ml). Calculated IC ₅₀ was 2.62 µg/ml. Cytotxicity assays and animal models: This experimentation was only carried out for <i>S. aureus</i> and <i>E. coli</i> .	Direct. OA inhibits the action hemolytic of PLY.	Pneumonia	A549 cells C57BL/6J mice	
[67]	Cholesterol (57-88-5)	Sterol (natural)	Hemolysis: PLY at a concentration of 500 ng, 100 ng, 50 ng, 10 ng, 5 ng, or 0 ng was mixed with 1% cholesterol. Cholesterol at 1% inhibited 50 ng PLY by 92%. Animal models: Pneumococcal keratitis was induced in rabbit corneas by intrastromal injection of <i>S. pneumoniae</i> or recombinant PLY (1µ). 25 h after infection, rabbit eyes were topically treated with a drop of 1% soluble cholesterol (40 mg). Seven ocular parameters were classified. Significant differences between cholesterol-treated corneas and untreated corneas were observed 48h post infection.	Direct. Cholesterol inhibits the PLY hemolysis. The hydrophobic region of domain 4 is responsible for interacting with cholesterol, specifically through a THR-LEU pair located in the L1 loop.	Keratitis	New Zealand White rabbits	D39
[93]	β-sitosterol (83-46-5)	Sterol (natural)	Cytotoxicity assays: The protective effects of β - sitosterol (BSS) on the toxicity of A549 cells were measured by the release of LDH showing that concentrations higher than 2 µg/ml protect the injury caused by 20 µg of PLY. Animal models: Mice infected were treated 1 h prior to the infection with BSS (80 mg/kg); a 70% survival rate at 5 days was observed.	Direct. β-sitosterol interacts with the toxin at THR459 and LEU460, competing with cellular cholesterol for binding to the toxin.	Pneumonia	A549 cells C57BL/6 mice	D39
[97]	Simvastatin (79902-63-9)	Statins (Synthetic)	Cytotoxicity assay: 1 µM simvastatin provided significant protection against higher doses of PLY, ranging from 0.4 to 1.6 µg/ml. 10 µM hydrophilic pravastatin also conferred significant protection against PLY. Animal models: 200 ng PLY was injected intratracheally in mice that have been injected intraperitoneally with simvastatin (20 mg/kg). Lungs of mice sacrificed 18 h later demonstrated lower injury and inflammation compared with the control group.	Indirect. Simvastatin significantly reduced the total cholesterol content of cells, but did not reduce the binding of PLY to cells. Simvastatin inhibits the cytotoxicity of PLY on airway epithelial cells.	Pneumonia	HBE1 ⁴ , NHBE ⁵ , A549 cells C57BL/6 mice	
[99]	Simvastatin (79902-63-9)	Statins (Synthetic)	Cytotoxicity assay: Cells were preincubated overnight with 0, 0.1, 1, or 10 µM simvastatin and incubated with PLY 3 µg/ml. The protection effect was observed at 1 µM. Animal models: WT and sickle cell disease (SCD) mice were treated with simvastatin (1 µg/g via intraperitoneal) daily for 5 days prior to bacterial challenge. Simvastatin had no significant effect on the survival of infected WT mice whereas the time of death was significantly delayed in SCD-treated mice.	Indirect. Statin treatment reduced platelet-activating factor receptor (PAFr) expression. PAFr binds to bacteria and is engulfed in vacuoles, invading the host cell.	Pneumonia and sepsis	HBMEC ⁶ C57BL/J6 mice	D39
[109]	Docosahexaenoic acid (6217-54-5)	Fatty acids Omega3 (natural)	Hemolytic activity: Percentages of hemolysis observed with PLY at 8.37 ng/ml alone or in the presence of 5 mg/ml Docosahexaenoic acid (DHA) were $56.3 \pm 2\%$ and $44 \pm 1.4\%$, respectively. For PLY at 4.19 ng/ml in the presence of the same DHA concentration, the percentages of hemolysis were $33.3 \pm 1.7\%$ and $22.7 \pm 1.3\%$, respectively.	Direct. DHA interferes with the binding of PLY to target cells. PLY-mediated influx of Ca2+, activation of NFxB (nuclear factor kappa-light-chain- enhancer of activated B cells) and IL-8 in neutrophils.		Human neutrophils	
[116]	9-(6-phenyl-2-oxohex-3-yl)-2- (3,4-dimethoxybenzyl)-purin- 6one (190666-14-9)	Purin-6-ones (Synthetic)	Transcellular electrical resistance (TER): HUVEC monolayers were incubated with PDP (1 and 0.01 µM) followed by toxin exposition. PLY (0.1 µg) evoked a strong decrease of TER, indicating increased endothelial cell permeability. PDP preincubation concentration dependently reduced the PLY-evoked increase in permeability. Animal models: Intranasal pneumococci-infected mice were treated 1 hour before infection with hydroxy-PDP (61.93 ±11.77 nM concentration final) infused subcutaneously. 48h after infection, lung hyperpermeability was decreased; however, inflammatory cell number did not change.	Indirect. Phosphodiesterase 2 (PDE2) inhibition (with PDP or hydroxy-PDP) decreased PLY induced human endothelial cell and alveolo-capillary membrane permeability. Protein expression of PDE2 in lung homogenates of pneumococci-infected lungs was significantly increased 48 h after infection. PDE2 inhibition decreased lung hyperpermeability induced by PLY.	Pneumonia	HUVEC ⁷ C57BJ/6 mice	ST3

Reference	Molecule (No CAS)	Molecule type	Toxicity assays	Action mechanism	Infection type	Cellular lines and laboratory animals	S. <i>pneumoniae</i> strain
[118]	CysLT1 antagonists Montelukast (158966-92-8)	Drug (Synthetic)	Animal models: Among BLT2-knockout mice intratracheally injected with 50 ng of PLY, 80% survived compared with the control group. Montelukast (5 mg/kg) improves PLY-induced (50 ng) acute lung injury in mice.	Indirect. BLT2 is a G protein- coupled receptor for leukotriene B4 and 12-HHT is a natural ligand that protects mice from lung injury caused by PLY. BLT2 suppress CysLT1 (receptor of cysLTs) expression. PLY triggers the production of cysteinyl leukotrienes (cysLTs) that activate CysLT1 expressed in vascular endothelial cells and bronchial smooth muscle cells, leading to lethal vascular leakage and bronchoconstriction. Montelukast is a CysLT1-selective antagonist that reverses the effects of PLY: vascular leakage and bronchoconstriction.	Pneumonia	BALB/c mice	
[121]	Clarithromycin (81103-11-9)	Antibiotics– Macrolide (Synthetic)	Animal models: Mice were intratracheally infected with <i>S. pneumoniae</i> . Clarithromycin (150 mg/kg) and Erithromycin (150 mg/kg) were administered orally 150 mg/kg every 12 h. 24h post infection, arterial oxygen saturation was improved whereas neutrophil number, IL-6 and PLY in the BALF decreased as compared to the control.	Unknown. Clarithromycin significantly downregulates <i>ply</i> gene transcription compared with ERY and the control.	Pneumonia	BALB/c mice	NU4471 Macrolide resistant D39
[122]	Ceftriaxone (104376-79-6) Levofloxacin (100986-85-4)	Antibiotics- Cephalosporin (Synthetic) Antibiotics- Quinolone (Synthetic)	Animal models: Mice were infected by intranasal challenge with multidrug-resistant <i>S. pneumoniae</i> and treated with intravenous doses of either Levofloxacin (LVX) (150 mg/kg) or Ceftriaxone (CRO) (50 mg/kg of body weight) or administered together. The percentage of survival at 72h was: LVX, 55%; CRO, 38%; and LVX+CRO 95% survival.	Unknown . Downregulating the expression of <i>ply</i> gene.	Pneumonia	BALB/c mice	Clinical isolate
[69]	C-terminal 70 amino acids of PLY (C70PLY)	Peptide	Cytotoxicity assays: HUVECs were treated for 24 h with PLY and C70PLY peptide at concentrations of 1, 10, 100, or 1000 nM. For PLY, cell viability was strongly diminished to 100 and 1000 nM (more than 40%) whereas no cell toxicity was observed for any C70 peptide concentration. Animal models: The anti-inflammatory potential of C70PLY peptide was determined in a model of atherosclerosis-induced rats. Rats treated with C70PLY reduced the formation of neointima which developed into atherosclerotic plaque in the control group.	Direct. PLY is a toll-like receptor (TLR) 4 ligand. TLR4 forms a complex with its specific coreceptor myeloid differentiation factor 2 (MD2). The peptide binds M2 inhibit toxin-induced lysis of host cells.	Atherosclerosis	HUVEC and PMN ⁸ Sprague–Dawley rats	
[64]	DM3 peptide: GLFDIWKWWRWRR-NH2 Indolicidin peptide derivative: ILAWKWAWWAWRR-NH2	Synthetic hybrid peptides	Animal models: Two models were assayed: systemic and pneumonia pneumococcal infection. Several combinations of peptide and penicillin were tested. In the systemic infection model, the best combination obtained (100% survival) were observed in mice treated with DM3 (20mg/kg) + penicillin (20mg/kg) treatment. In the pneumonia infection model mice survival was not observed in any treatment.	Direct. DM3 has a strong affinity PLY (interaction with ALA370, TYR371, TYR376, ASN400, ASP403, CYS428, ALA432, TRP435, TRP436 of PLY) and showed therapeutic synergism in combination with penicillin. Indolicidin peptide derivative bounded better than DM3 (In silico).	Systemic and pneumonia	ICR mice	Penicillin Resistant strain from clinical isolate
[7]	Mannose receptor peptides (MRC-1)	Peptide	Cytotoxicity assays: Human THP-1 macrophages treated with PLY (0.5 µg/ml) and 100 µM peptides. Significant reduction of toxicity was observed (~ 50%) with P2 and P3. Animal models: Two models were assayed: mice pneumonia infection and zebrafish fertilized embryo infection. Mice were infected intranasally with bacteria mixed with peptide P2 alone (5 µg) or P2 conjugated to CaP NPs (5 µg peptide; 25 µg CaP NPs) (biocompatible calcium phosphate (CaP) nanoparticles (NPs) as peptide nanocarriers). Percentage of survival at 3 days were increased to 50% in mice treated with P2-NP and 20% at 2 days in mice treated with P2. Infected zebrafish fertilized embryos were microinjected into the yolk sac with 500 CFU mixed with P2 or P2-conjugated NPs (1nM). After 100h post-infection, only 50% of embryos survived the infection versus 80% of embryos treated with P2	Direct. The cholesterol binding loop of PLY (W433, W435, W436, E434) bind the C-type lectin domain 4 (CTLD4) of MRC-1. Peptides competitively inhibit the PLY binding.	Pneumonia and systemic	Human THP-1 macrophages C57BL/6 mice Zebrafish infection model	TIGR4 D39

Reference	Molecule (No CAS)	Molecule type	Toxicity assays	Action mechanism	Infection type	Cellular lines and laboratory animals	S. pneumoniae strain
[37]	TNF-derived tonoplast intrinsic protein (TIP peptide). PKC-α inhibitor Ro 32–0432 hydrochloride (Bisindolylmaleimide XI hydrochloride) (145333-02-4)	Peptides and drug (Synthetic)	Measurement of Transendothelial Electrical Resistance (TER): PLY treatment of human lung microvascular endothelial cells induces a dose- dependent increase in intracellular Ca2 + concentrations. Pretreatment of the cells for 1 hour with PKC-α inhibitor Ro32-4032 (10 nM) or with TIP peptide (27 µM) significantly reduces this effect. Animal models: Intratracheal instillation of 3.125 µg PLY/kg induces a significantly increased pulmonary endothelial permeability. Cotreatment of the mice with the PKC-α inhibitor Ro32-0432 (49 µg/kg, intratracheally) or with the TIP peptide (2.5 mg/kg, intratracheally) significantly inhibits PLY-induced capillary leak.	Indirect. PLY causes increased arginase activity and PKC- α activation, both involved in endothelial dysfunction. A specific PKC- α inhibitor (Ro 32– 0432) and the TIP peptide inhibit PKC- α and arginase which respectively mitigates the endothelial hyperpermeability.	Pneumonia	HLMVEC ⁹ and HPAEC ¹⁰ C57BL/6 mice	
[129]	GHRH agonist: JI-34 peptide	Peptide analogs	$\label{eq:meansature} \begin{array}{l} \mbox{Measurement of Transendothelial Electrical} \\ \mbox{Resistance (TER): PLY treatment of H441 cells (30 ng/mL) leads to a significant reduction of both inward and outward currents. The subsequent treatment of the cells with JI-34 (1 \muM) restores the Na+ current. \\ \mbox{Animal models: JI-34 (100 µg/kg) completely inhibits the increase in lung wet-to-dry weight ratio (edema) caused by PLY instillation (6.125 µg/kg). \end{array}$	Indirect. JI-34 can restore Na + uptake and blunts phosphorylation of MLC (myosin light chain) and VE-cadherin induced by PLY. It also mitigates the endothelial hyperpermeability.	Pneumonia	H441 cells ¹¹ and HLMVEC C57BL6 mice	
[130]	Vasculotide	Peptide modified	Measurement of Transendothelial Electrical Resistance (TER): Cells were pretreated with Vasculotide (VT) (2, 10, and 50 ng/ml) and then with PLY (0.75 µg/ml). Preincubation with VT at 50 ng/ml attenuated the PLY-induced TER. Animal models: Mice were transnasally inoculated with bacteria a nd i.v. with VT (100, 200, or 500 ng). VT 500 ng significantly reduced edema formation but leukocyte recruitment and cytokine production 48h post-infection were not affected.	Indirect. Angiopoietins are regulators of inflammation and vascular leakage and ligands for the receptor tyrosine kinase Tie2. VT bind Tie2 mimic Angiopoietin-1.	Pneumonia	Murine lung endothelial cells (mLEC) C57BL/6 N	NCTC7978
[133]	Z-VAL-ALA-OH (24787-89-1)	Peptide modified	Cytotoxicity assays: HUVEC were pre-incubated with 50 µg/ml zVAD for 30 minutes and then stimulated with <i>S. pneumoniae</i> R6x. zVAD completely inhibited pneumococci-related DNA fragmentation (apoptosis). Pneumolysin-deficient mutant R6xΔply induced apoptosis less effectively than R6x.	Indirect. Deletion of the gene coding for PLY reduced pneumococci-induced apoptosis in HUVEC, involving mitochondrial death pathways. Programmed cell death could be strongly reduced by pan-caspase inhibitor zVAD.		HPMEC-ST1.6R ¹² and HUVEC	R6x
[31]	zVAD-fmk (187389-52-2)	Peptide modified	Animal models: Mice were intranasally inoculated with strain WU2 and zVAD-fmk (20 µg) every 12h. Percentage of survival was 50% in treated mice versus 90% in the control. This treatment diminished survival rate.	Indirect. PLY is a ligand for the TLR-4 receptor, activating the innate immune response. TLR- induced apoptosis is caspase 3 dependent and is a defense mechanism against infection. Thus, inhibition with zVAD-fmk decreases survival to infection.	Sepsis	C3H/HeOuJ and C3H/HeJ mice	WU2 and A66.1 Xen 10
[66]	Mg ²⁺	Cation	Cytotoxicity assay: Mouse glial cells, 4 HU/ml PLY and 2 mM Mg treatment, diminished LDH release at 60 min after PLY challenge. Animal models: Two models of induced meningitis were assayed in mice and infant rats. Infant SD rats were intracerebroventricular (i.c.v.) injected with PLY (4 HU/ml) and i.p. with MgCl2 (500 mg/kg) which significantly reduced brain edema. Mice received an intracerebral (i.c.) injection of 1000 CFU and with three i.p. doses MgCl ₂ 30.45 mg/ml prolonged survival at 36 h; mortality was 2/ 18 in the treated group versus 8/19 in the control group.	Unknown. Magnesium diminishes interstitial brain edema caused by PLY. Mg2 + diminishes the pore-forming capacity of PLY without inhibit toxin binding to cells.	Meningitis	Primary glial cultures C57BL/6 mice SD rats	D39
[135]	Zn ²⁺	Cation	Cytotoxicity assay: Cells were incubated with PLY lng/ μ l. The protective effect of zinc at 1 μ M was 39.3% and 50.22% for IHCs and OHCs, respectively. At 300 μ M of zinc, protection was significantly increased with 62% and 55.2% for IHCs and OHCs, respectively.	Unknown . Zinc inhibits the incorporation of PLY into the membrane.		Cochlear hair cells (HCs) and Inner hair cells (IHCs) Wistar rats	

Reference	Molecule (No CAS)	Molecule type	Toxicity assays	Action mechanism	Infection type	Cellular lines and laboratory animals	S. pneumoniae strain
[139]	Ca ²⁺	Cation	Cytotoxicity assay: Cells were incubated with PLY (0.5–0.025 μg/ml) and Ca 2mM or Ca-free. Minimal toxicity was observed in presence of 2mM Ca whereas in Ca-free buffer, high lytic capacity of PLY was observed even at sublytic concentrations.	Unknown. Reduction Ca concentrations improve the PLY membrane binding. Although the mechanism is not fully understood, it could be related to membrane fluidity at different Ca concentrations rather than to a direct binding of the cation to PLY.		Primary astrocytes from C57BL/6 mice	
[140]	Ca ²⁺	Cation	$\label{eq:constraint} \begin{array}{l} \hline Cytotoxicity assay: A549 cells treated with 0.1 \mug/ mL PLY induces cell death in approximately 50% of the cells after 18 hours as revealed by 2 cellular phenotypes showing either cell death or survival. The surviving population cleared the [Ca2+]m significantly faster than the dying population. Treatment of PLY-induced cells with DMSO reversed the effect, while treatment with the benzothiazepine compound CGP-37 157 (10 \muM) (a known Na/Ca exchanger inhibitor) resulted in a decreased number of surviving cells. $	Indirect. Some alveolar epithelial cells survive apoptosis induction after PLY challenge. Mitochondrial calcium flux is decisive for surviving PLY stimulation.		A549 cells	
[141]	Antibodies	Protein	Animal models: Two animal models were used, toxin lethality and pneumonia. Toxin lethality model determined the $LD_{50} = 1.06 \ \mu g$ of PLY. 1 h before intranasal infection with <i>S. pneumoniae</i> , 100 μg of anti-PLY monoclonal antibodies were intravenously injected. Media survival times was recorded 20 days after infection. The survival rate of mice that received a mix of three monoclonals was 10/20 versus a survival rate of 2/20 in the control group.	Direct. Anti-PLY antibodies increased median survival times and survival rates, decreased bacteria number, and leukocytes infiltration.	Pneumonia	MF-1 mice	D39
[68]	Antibodies	Protein	Animal models: In slit lamp examinations (SLEs) seven ocular parameters were graded. At 36 and 48 hours after infection by intrastromal injection with <i>S. pneumoniae</i> , SLEs were significantly lower in rabbits passively immunized (ear vei nijection) with PLY antiserum (IgG titer of 409,600).	Direct. Anti-PLY serum elicited protection against severe corneal opacity and massive infiltration of PMNs.	Keratitis	New Zealand White rabbits	WU2
[70]	Antibodies	Protein	Animal models: After intranasal inoculation of bacteria to establish nasopharynx colonization, 10 µg or 25 µg of anti-PLY antibodies were administered intravenously. 3 days after inoculation, there was a significant decrease in CFU colonization in mice passively receiving anti-PLY IgG (25 µg).	Direct. Anti-PLY antibodies reduce adherence of bacteria to epithelium in vivo (but not in vitro) possibly due to the role of PLY in biofilm formation.	Nasopharyngeal colonization	A549 cells C57BL/6 mice	TIGR4 RX1

¹A549 cells: Human lung epithelial cells

²Hep2: Human larynx carcinoma epithelial cells (ATCC CCL-23)

³RAW264.7 cells: Macrophage-like, Abelson leukemia virus-transformed cell line derived from BALB/c mice

⁴HBE1

⁵NHBE

¹¹H441 cells: Human Bronchial Epithelial cells

⁶HBMEC: Human Brain Microvascular Endothelial Cells

⁷HUVEC: Human Umbilical Vein Endothelial Cells

⁸PMN: Polymorphonuclear Neutrophils

⁹HLMVEC: Human Lung Microvascular Endothelial Cells

¹⁰HPAEC: Human lung pulmonary artery endothelial cells

¹²HPMEC-ST1.6R: Pulmonary endothelial cell line.

https://doi.org/10.1371/journal.pone.0282970.t001

Contrastingly, compared to the total number of trials, there is a high number of antibiotics studies in the context of pneumonia.

Types of molecules and drugs tested

Plant-derived compounds: Polyphenols, flavonoids, tannins, quinones, phenethylamines and terpenoids. The seventeen plant-derived compounds included in this review belonged to six categories according to their chemical characteristics. Some of them present antibacterial activity [71, 72] and therapeutic effects on acute lung injury since regulating the TLR4/NF- κ B signaling pathway, NLRP3 inflammasome activation and the MAPK signaling pathway [73, 74].

Polyphenols are a large and diverse family of naturally occurring organic compounds abundant in plants. They are generally subdivided into phenylpropanoids, flavonoids, hydrolysable tannins, and condensed tannins. In this work, flavonoids and tannins are described independently for greater clarity. An example of polyphenol is Verbascoside, a phenylpropanoid that binds to the cleft between domains 3 and 4 of PLY (Asp471, Asn470, Glu277, Tyr358, and Arg359), as was shown by the molecular dynamic's simulation, thus blocking the conformational transition from monomeric to oligomeric form and inhibiting its lytic activity. Verbascoside was tested in animals suffering from pneumonia with protective effects and reduction of levels of TNF- α and IL-1 β [75]. Flavonoids possess the basic structure of a chromone (1,4-benzopyrone) moiety connected to a phenyl ring at position 2. Some examples such as Apigenin [76], Acacetin [77], Amentoflavone [78], and Morin [79] decreased the hemolytic activity of PLY by inhibiting oligomerization. In animal models, the inflammatory cytokines INF- γ and IL-1 β in the lungs of mice treated with these compounds decreased significantly. Other flavonoids such as Epigallocatechin gallate (EGCG) significantly increased the survival of infected animals due to it affected PLY oligomerization by binding to the toxin residues Ser256, Glu277, Tyr358, and Arg359 [80]. In addition, this molecule interferes with the biofilm formation and bacterial adherence to cells. Another interesting flavonoid is quercetin, which has been tested in animal models of lung infection, with survival rates of 80% compared to 60% in untreated mice [81]. For other flavonoids such as Dryocrassin ABBA, data from animal models are not available, but in vitro results showed relative bactericidal activity and the ability to interfere with toxin oligomerization capacity at concentrations ranging from $0.5 \,\mu\text{g/ml}$ to 2 µg/ml [82]. Recently, 27 hydrolysable tannins were tested by Maatsola et al. (2020) [83]. Nanomolar concentrations of Pentagalloylglucose (PGG) and Gemin A were capable of inhibiting the PLY cytolytic capacity. Molecular modeling suggests that PGG also binds to the cleft between domains 2, 3 and 4 (Glu42, Ser256, Asp257, Glu277, and Arg359).

Quinones are a class of organic compounds formally derived from aromatic compounds. Shikonin [84] and Juglone [85] are quinones (naphthoquinones) that inhibited pore formation by means of interfered toxin oligomerization. Aloe-Emodin is a quinone (anthraquinone) with antifungal activity. The authors have proposed this compound together with photodynamic therapy to treat superficial infections of antibiotic-resistant gram-positive bacteria (*Enterococcus faecalis, Staphylococcus aureus*, and *S. pneumoniae*). Irradiation generates reactive oxygen species (ROS), leading to the destruction of biomolecules and the killing of bacterial cells; however, the mechanism by which cytotoxins expression (such as PLY) decreases is unknown [71]. Application *in vivo* requires future investigation.

Ephedrine hydrochloride and Pseudoephedrine hydrochloride are phenethylamines from *Ephedra sinica* granules [86] and MXSGT (Ma-xing-shi-gan-tang) [87] are compounds obtained from plants used in traditional Chinese medicine. All of them seem to inhibit the oligomerization of the toxin, although more studies are necessary to specify which residues of PLY are involved. In addition, Ephedrine hydrochloride is part of the WHO model list of essential medicines and is used to prevent low blood pressure during anesthesia, and for the treatment or prevention of attacks of bronchospasm in asthma. It is a very active agonist adrenergic on the receptors of the sympathetic nervous system. Ephedrine hydrochloride increased anti-inflammatory cytokine production IL-10 and decreased the production of pro-inflammatory cytokines TNF-α and IL-12 in dendritic cells in *Staphylococcus aureus*-induced

peritonitis animal models [88]. Current clinical trials are focused on the effect of this compound on sinusitis, rhinitis, etc.

Oleanolic Acid, Betulin and Hederagenin are terpenoids. Oleanolic Acid and its analogues significantly inhibited the activity of important β -lactamases like NDM-1, KPC-2, VIM-1, and OXA-1 as well as the hemolytic action of cytolysins like PLY [89]. For *S. aureus* α -hemolysin, the authors showed through molecular modeling how oleanolic acid can interfere with toxin oligomerization and protect *S. aureus*-infected mice in a combination therapy with β -lactams. More research is needed to see these effects *in vivo* on other toxins, including PLY. Betulin [90] and Hederagenin [91] are triterpenes extracted from plants that interfere with the PLY oligomerization process, without toxic effects against epithelial cells, but have not yet been tested in animal models of pneumococcal infection.

Although most of these above mentioned molecules inhibit or hinder the PLY oligomerization, only three articles [75, 78, 85] studied potential interactions between molecules and toxin by using molecular modelling and docking calculation. Thus, it was probed that the flavonoid Amentoflavone was able to interact with domain 2 (Arg359) and domain 3 (Ser254, Glu277) of the toxin; similarly, to the polyphenol Epigallocatechin gallate, which forms strong interactions with the PLY domain 3 (Ser256, Glu277) and domain 2 (Tyr358, Arg359). Furthermore, Asp 471 from domain 4 of PLY could form a strong hydrogen bond with the hydroxyl group of the benzene ring on the right side of the polyphenol Verbascoside. On the other hand, the interactions of the hydrolysable tannin PGG and PLY take place through flexible galloyl groups of PGG and the pocket formed by domains 2, 3 and 4 of PLY [83]. To our knowledge, no controlled clinical trials have addressed the issue of adjuvant plant-derived compound therapy in the clinical setting of pneumococcal infections. As exception, recent clinical trials using Quercetin (or products containing it) for treating pneumonia in patients with COVID-19 have showed a significantly reduced length of hospitalization, noninvasive oxygen therapy, and number of deaths [92], which clearly confirms its potential as an anti-inflammatory molecule.

Sterols. Two sterols, cholesterol and β -sitosterol, inhibited the binding of the toxin to the membrane. Cholesterol is the natural ligand for PLY in the eukaryotic cell membrane. Marquart et al. (2007) [67] showed that cholesterol has a bactericidal effect both *in vitro* and in corneas of rabbits with experimental pneumococcal keratitis. The authors suggest that cholesterol would not only bind to PLY, but also to bacteria and kill them. β -sitosterol is a phytosterol with chemical structure very similar to cholesterol, that has been assayed in murine pneumonia model with good protection results [93]. Additionally, β -sitosterol possess anti-inflammatory properties, causes a dose-dependent inhibition of IL-6 and TNF- α in endotoxin-activated human monocytes, and has beneficial effects on the immune system by increasing the number of viable peripheral blood mononuclear cells and activating the dendritic cells [94, 95]. A detailed study of the binding between five sterols and PLY has confirmed that PLY interacts with these molecules through residues Tyr371, Val372, Leu460, and Tyr461. In the five natural sterols studied, the critical structure is the C22-C23-C24-C25 carbon bonds [58]. We did not find clinical trials of pneumonia related to these sterols.

Statins. The protective effect of simvastatin and pravastatin against the cytotoxic effect of PLY *in vitro* was described by Statt et al. (2015) [96, 97]. Oral treatment with simvastatin in pneumonia mice reduced neutrophil infiltration, maintained vascular integrity, and decreased chemokine production [98]. Statins may trigger mevalonate-independent pathways partially via a calcium increase and p38 MAPK activation. Statins act as competitive inhibitors of HMG-CoA reductase (HMGCR), the rate-limiting enzyme of cholesterol synthesis. Simvastatin was also used in a mouse model of sickle cell disease, which is characterized by hemolytic anemia and chronic inflammation, and was shown to lead to a high incidence of invasive pneumococcal pneumonia. The last was due to upregulation of platelet-activating factor

receptor (PAFr). In this context, Simvastatin is useful as it reduced PAFr expression and interfered with toxin pore formation and host cell bacteria invasion [99]. Statins are widely used in the treatment of cardiovascular disease in humans. In last years, statins have begun to be studied in the context of infectious diseases like tuberculosis, AIDS, and COVID-19 [100–102]. Several clinical trials have been carried out with simvastatin in patients with CAP with contradictory results. Some authors find that prior use of this statin improves mortality in patients admitted with CAP [103] while other authors find no difference [104–106]. On the other hand, the administration of 20 mg in individuals admitted with CAP does not show differences in cytokine levels [107]. Recently, clinical trial have been carried out in older adults with derivate CAP bacteremia with encouraging results [108]. Simvastatin could be a good candidate to complement antibiotic therapy. In addition to the bactericidal effect against Gram positive microorganisms, statins have pleiotropic effects on cells of the immune system (i.e., stimulate autophagy in macrophages, increase the number of NK and NKT cells, inhibit MHC-II expression, increase serum levels of IL-10) [108].

Omega-3 fatty acids. Omega-3 fatty acids are polyunsaturated fatty acids (PUFAs) characterized by the presence of a double bond, three atoms away from the terminal methyl group in their chemical structure. The structure of docosahexaenoic acid (DHA) is a carboxylic acid with a 22-carbon chain and six *cis* double bonds, with the first double bond located at the third carbon of the omega end. DHA decreased IL-8 PLY mediated in human neutrophils by interfering with Ca²⁺ influx [109]. Also, DHA hinder the binding of toxin to cellular membrane of neutrophils by an unknown mechanism. However, it is accepted that lipid rafts are putative binding sites for cytolysins and that omega-3 polyunsaturated fatty acids exclude proteins from these lipid rafts in eukaryotic cell membranes.

Omega-3 fatty acids are widely known in the field of nutrition and their role is being investigated in neurodegenerative diseases, sleep, etc. As molecules regulating inflammation in the context of infectious diseases, they have been reviewed by Sandhaus & Swick (2021) [110]. The specialized pro-resolving mediators (SPMs) are enzymatically derived from essential fatty acids, including arachidonic acid, eicosapentaenoic acid, and DHA, and have important roles in the resolution of tissue inflammation. DHA is converted to D-series resolvins, protectins, and maresins via 12- and 15-LOX enzymes. DHA-derived SPMs have been shown to be essential for modulating the number and response of T-helper cells to resolve chronic inflammation [111]. Exogenous administration of SPMs in infectious conditions has been shown to be effective at improving infection clearance and survival in preclinical models [112]. Clinical trials carried out with omega-3 fatty acids in the treatment of acute lung injury and acute exacerbations of chronic respiratory disease, asthma, and pneumonia [113, 114], did not show the expected improvement results. However, in patients with COVID-19, C-reactive protein levels, erythrocyte sedimentation rate, and some clinical symptoms were reduced [115].

Purin-6-ones. PLY has a known role in increasing the permeability of fluids from microcapillaries to alveolar spaces. Witzenrath et al. (2009) [116] demonstrated the involvement of phosphodiesterase 2 (PDE2) in the hyperpermeability of lung tissue during pneumococcal pneumonia. Purin-6-ones, 9-(6-phenyl-2-oxohex-3-yl)-2-(3,4-dimethoxybenzyl)-purin-6-one (PDP), and hydroxy-PDP are specific inhibitors of PDE2. PDP decreased PLY-induced permeability *in vitro* in endothelial cell cultures and *in vivo* in murine pneumococcal pneumonia models, improving lung damage without interfering with the recruitment of immune system cells into lung tissue. PDE2 enzymes have been also found in various tissues and cells, including pulmonary arterial smooth muscle cells, endothelial cells, platelets, and macrophages. Up to now, there have been no studies available on the use of PDE2 inhibitors in the clinical studies carried out on patients with pneumococcal pneumonia. However, nonselective inhibitors of PDEs as Theophylline has been used in the treatment of bronchial asthma and chronic obstructive pulmonary disease (COPD) for more than 50 years. Various (selective) PDE3, PDE4, and PDE5 inhibitors have also demonstrated stabilization of the pulmonary epithelialendothelial barrier and reduction the sepsis- and inflammation-increased microvascular permeability, and suppression of the production of inflammatory mediators, which finally resulted in improved oxygenation and ventilatory parameters [117].

Thioethers. Montelukast is a synthetic thioether that acts by decreasing the hyperpermeability and bronchoconstriction produced by the toxin in the lungs [118]. PLY triggers the production of cysteinyl leukotrienes and their receptor CysLT1 in bronchial smooth muscle cells; the interaction of vascular endothelial cells increased vascular permeability, edema, influx of eosinophils, and neutrophils. Montelukast is a CysLT1 antagonist that increases survival in mice with induced pneumococcal pneumonia. In addition, this compound is frequently used as a treatment in patients with asthma and allergies. Recently, montelukast treatment for pneumonia, caused by *Mycoplasma pneumoniae*, and Sars-CoV-2 decreased inflammation markers and contributed to improvements in respiratory insufficiency [119, 120].

Antibiotics. Clarithromycin is a derivative of erythromycin and differs structurally in the substitution of an O-methyl group for the hydroxy group at position 6 of the lactone ring. Clarithromycin negatively regulates *ply* gene transcription and activates pneumococcal autolysis [121]. On the other hand, in animal models of pneumonia caused by multidrug-resistant S. pneumoniae, the therapy combining levofloxacin and ceftriaxone modulated the inflammatory response (decreased number of neutrophils and polymorphonuclear neutrophils and vascular leakage; decreased nitric oxide production) of pneumococcal infection at sub-MIC levels by downregulating virulence genes (ply and lvtA), cyclooxygenase-2 (COX-2), and inducible nitric oxide synthase (iNOS) [122]. Generally, macrolides and macrolide-like agents have antiinflammatory activities, including modulation of cytokine production [123]. Other antibiotics with anti-inflammatory properties are quinolones such as levofloxacin, due to its ability to join to the hydrophobic region of the TLR4-MD2 complex [124]. Clarithromycin is used in the treatment of pneumococcal infections in combination with β -lactams and in some clinical trials using clarithromycin as treatment (revised by Anderson & Feldman, (2017) [59]); the results supported its beneficial role in the anti-inflammatory/immunomodulatory activities. More recently, in a clinical trial in patients with COVID-19, it appears that it could be beneficial for early control of fever and early negative conversion of PCR [125]. However, increasing macrolide resistance in some CAP ethological agents (S. pneumoniae and M. pneumoniae) and its effect on decreasing the natural T-cell immune response [126], is contributing to reconsideration of their use in the future.

Peptides. In this review, seven peptides are presented as candidates for adjuvant therapy. Three of the peptides [64, 65, 69] (J34 binds to the SV1 splice variant of the GHRH receptor, TIP binds PKC- α , and vasculotide binds to the Tie2 angiopeptin receptor) act on endothelium hyperpermeability thus minimizing fluid extravasation into alveolar space. Le et al. (2015) [64] showed how synthetic hybrid peptides, with antimicrobial activity improved survival and reduced the sequelae of lethal pneumococcal systemic infection in animal models. Molecular docking analysis showed that one of the peptides (DM3) had a strong affinity for PLY, also for autolysin and pneumococcal surface protein A (pspA). DM3, in combination with penicillin, showed the recovery of all the treated septicemic animals, but it was not effective in pneumonia, therefore more studies are necessary (Le et al., 2015) [64]. Actually, antimicrobial peptides account for a promising alternative to conventional antibiotics since they can be designed to act on various targets and are less likely to generate resistance [127]. The problems encountered for its clinical application are related to its low absorption and toxicity to human cells. In most cases, these drawbacks can be solved with chemical modifications and *in silico* design [128].

The growth hormone-releasing hormone (GHRH) agonist, JI-34, decreased PLY-mediated endothelial hyperpermeability by reducing the phosphorylation of myosin light chain and vascular endothelial (VE)-cadherin and restoring basal levels of Na⁺ [129]. Another example is Vasculotide (a tetrameric peptide with four octapeptides (NH₃-CHHHRHSF-COOH), which are covalently attached through NH₂-terminal maleimide) which improved the barrier function of the endothelial epithelium during pneumococcal pneumonia (Gutbier et al., 2017) [130]. The action was mediated by its binding to the Tie2 receptor of angiopeptin (Ang/Tie) thereby protecting against increased levels of fluid invading the intra-alveolar space and further pulmonary edema. Other drugs targeting the Ang/Tie pathway are being tested for the control of pulmonary vascular disorders in clinical trials. AV-001, a Vasculotide-derived peptide (Vasomune Therapeutics) is currently in phase I. The results showed that AV-001 is safe and well tolerated in patients with COVID-19, acute respiratory distress syndrome (ARDS), or COVID-19-associated ARDS [131].

The TIP peptide (TNF-derived tonoplast intrinsic protein) and the Ro 32–0432 peptide (bisindolylmaleimide XI hydrochloride) are Protein Kinase C- α (PKC- α) inhibitors. PKC- α is stimulated by the Ca²⁺ influx mediated through PLY-induced pores which increased RhoA / RKK; arginase I reduces NO generation *in vivo* and consequently, microvessel leakage. TIP peptide diminished PKC- α activation and therefore, reduced PLY-induced endothelial hyper-permeability [129]. AP301 (Solnatide) is a circularized derivative of TIP peptide and has a Phase IIB clinical trial in ARDS [132] and is also being tested in patients with COVID [132]. The expected results, based on the previous studies, are activation of the pulmonary sodium ion channel (ENaC) to directly activate alveolar liquid clearance and reduce the leakage of blood and fluids from the capillaries in the airspace, i.e., accelerate the resolution of alveolar edema and reduce barrier injury in the lung.

Among the therapeutic peptides studied, some of them were designed to form hydrogen bridges with the cholesterol-interaction loop of different CDCs (PLY, LLO, and SLO) in a way that inhibits their hemolysis capacity and their pro-inflammatory activity. Thus, the named MRC-1 peptides bind PLY in domain 4 and inhibit binding to the human mannose receptor (MRC-1/CD206, a phagocytic receptor for bacteria, fungi, and other pathogens) [65]. Therefore, they reduced PLY-induced damage to the integrity of the epithelial barrier, the release of the chemokine IL-8, and the pro-inflammatory cytokines TNF- α and IL-12 as well as blocked bacterial invasion into the epithelium. Another example includes a peptide generated with the 70 C-terminal amino acids of PLY (C70PLY), which competed with MD2 to bind to TLR-4, inhibiting the effect of lipopolysaccharide on the phosphorylation of the ERK1/2 and NF κ B-p65 subunit. TLR-4 is a member of the toll-like receptor family that senses pathogen-associated molecular patterns, stimulating cytokine production and apoptosis. Also, its activation leads to an intracellular signaling pathway NF- κ B and the activation of the innate immune system [69].

As mentioned above, the innate immune system recognizes the pathogen components through TLR receptors, which trigger the host's first defense system. The inflammatory action of PLY is mediated by TLR-4 and induces caspase 3-dependent apoptosis in macrophages. zVAD (carbobenzoxy-valyl-alanyl-aspartyl-[O-methyl]-fluoromethylketone) is a modified peptide caspase inhibitor used to block apoptosis [31, 133]. However, apoptosis appears to be a defense mechanism against invasion by pneumococcus; therefore, inhibition of apoptosis results in increased mortality in animal models of pneumococcal pneumonia. Yet, cells can recover from PLY-induced apoptotic cell death by controlling mitochondrial $[Ca^{2+}]_m$ flux which contributes to the preservation of membrane potential ($\Delta\Psi$ m). PLY binding cholesterol in rich cellular membranes leads to large pores (250–350 Å) which contributes to a rapid influx of Ca²⁺ into the cytoplasm. Ca²⁺ induces repair mechanisms for protection of the host cell

[134]. One of them is sequestering Ca^{2+} into the mitochondria. However, exceeding the mitochondrial capacity can result into release of pro-apoptotic factors such as cytochrome C (cytC) or apoptosis inducing factor (AIF) [26]. We did not find clinical trials related to with the treatment of diseases with zVAD.

Cations. Cations like Ca^{2+} , Mg^{2+} , and Zn^{2+} hinder pore formation thereby decreasing the cytolytic capacity of PLY. Hupp et al. (2017) [66] showed that Mg²⁺ decreases the pore-forming capacity of PLY in primary glial cells, improving survival in animal models of meningitis. Cations are known to influence membrane fluidity or inhibit toxin binding, but these hypotheses did not hold for Mg⁺² since it does not prevent the binding of the toxin to the membrane, nor does it seem to change its characteristics. Another divalent cation, Zn²⁺ [135], protected rat cochlear hair cells from the toxic action of PLY by preventing its binding to the membrane. Recently, the deficiency of Zn^{2+} has been involved in a higher risk of acute infection in nasopharyngeal S. pneumoniae carriers. Therefore, supplementation with this element would improve immune system performance and consequently reduce pneumonia [136], Recent research on the relationship between micronutrients and some infections has shown improved immune function [137, 138]. Unlike the other cations, Wippel et al. (2011) [139], using brain tissue, postulated that Ca^{2+} could vary the fluidity of the membrane rather than having a direct action on the toxin because reductions in Ca²⁺ concentrations improve the binding of the PLY membrane thus increasing its lytic capacity. However, S. pneumoniae is capable of inducing apoptosis thanks to the pores formed by PLY which produce rapid inflows of mitochondrial calcium [Ca²⁺]_m thereby favoring fragmentation, loss of motility, and membrane potential in addition to activation of caspase 3. Nerlich et al., (2021) [140] showed that a significant number of alveolar epithelial cells survived caspase activation after being challenged with PLY, critically regulating $[Ca^{2+}]_m$ to control cell fate after attack by PLY. Therefore, calcium ions were postulated to be a useful candidate in therapeutic intervention during pneumococcal infection.

Antibodies. Antibodies against PLY have been used against pneumonia, keratitis, and nasopharyngeal colonization in animal models of infection [68, 70, 141]. The protective effect of mouse monoclonal antibodies in reducing PLY proinflammatory properties was clearly demonstrated as well as an increased survival rate with decreased bacteria lungs colonization, leukocyte infiltration, and lung injury [141]. Similarly, a reduction in toxin effects was observed in an induced keratitis model in rabbits using anti-PLY polyclonal antiserum [68]. Interestingly, the efficacy of human anti-PLY antibodies in reducing S. pneumoniae nasopharyngeal colonization was also demonstrated in mice [70]. Although PLY is an intracellular protein that is released during bacteria lysis, it would also be located on the cell surface, where it would play a role in the aggregation and formation of biofilms. Potential rejection reactions to repeated doses of antibodies have been solved through the humanization of monoclonal antibodies, which has facilitated their use for the treatment of various diseases [142]. This, together with the fact that immunoglobulins have been used since ancient times for the treatment of infectious diseases, makes this strategy very promising. In fact, the effect of PLY on platelets and erythrocytes can be neutralized by polyvalent human IgG and trimodulin [143]. In a clinical trial in patients with S. pneumoniae CAP, treatment with trimodulin (182.6 mg/kg, for 5 consecutive days) improved survival compared to patients treated with placebo [45]. Drugs developed against toxins from other microorganisms, including monoclonal antibodies, antibody fragments, antibody mimetics, have recently been reviewed [144]. It is important to highlight the phase I clinical trial using liposomes to capture S. pneumoniae toxins [145].

Network of compounds and related proteins. In order to evaluate the molecular interactions of the 39 compounds with neutralizing toxin properties described in the selected articles, the EXTRACT 2.0 tool was used for text mining of biomedical named entities and ontology terms such as diseases, subcellular localizations, tissues, drugs, and other small molecule



Fig 3. Knowledge network of the protective molecules against PLY. PLY interactions (edges) with host molecules (cyan circles), cells (cyan rectangles), and cellular processes (orange diamonds) is shown. Three types of connections are distinguished: the red arrows indicate inhibition (\perp) whereas the blue arrows refer to activation (\uparrow) and the grey arrows show interactions without inhibition or activation. The width of the lines reflects article ranking. The right panel represents the abundance and color of the molecules grouped by type. The network was built using Cytoscape software after text mining articles with EXTRACT 2.0.

https://doi.org/10.1371/journal.pone.0282970.g003

compounds [62]. With the resulting dataset, a knowledge network (Fig 3) was built using Cytoscape (see Materials and methods). Analysis of the network reveals that the most affected process is the formation of the pore with 19 interactions (edges), followed by hyperpermeability and inflammation (showing nine interactions), and membrane binding (seven interactions). The relevance and complexity of pneumonia is evidenced by the high number of molecules (24) used for its treatment while other diseases show three or fewer interactions with therapeutic molecules. Regarding host cell proteins, TNF- α was the protein most affected with 6 interactions, followed by IL-6 and, IL-1 β (both showing five interactions), and VE-cadherin (three interactions). The presented knowledge network visually shows the summary of

the reviewed articles. In addition to this, the structure of the network reveals the molecules with the greatest interactions which can be useful in the search for new treatments for the neutralization of the toxin. Drug repurposing identifies new uses of drugs outside their original medical scope, considering that, sometimes, one molecule can act on multiple targets [146]. Recently, a network medicine platform based on systems pharmacology was used to reposition antiviral drugs against SARS-CoV-2 [147]. This approach aims to reduce both development costs and the time it takes for already approved drugs to reach the market.

Current limitations and future perspectives

Despite the effective development of antibiotics and more recently pneumococcal vaccines, pneumococcal pneumonia continues to cause high morbidity, even though actually the mortality caused to invasive pneumococcal disease is declining due to childhood vaccination and herd protection [148]. The aging of the population and the combination of pneumococcal disease with other important viruses such as HIV, influenza and, more recently, SARS-CoV-2, could be behind these high levels of mortality and morbidity. To proceed in the discovery of new strategies to combat this disease, we decided to focus our work on PLY and its role in the performance of pneumococcal infections.

The combination of antibiotics with other molecules that can somehow block the action of PLY, was presented as a hopeful therapy aimed at significantly reducing the severity of pneumococcal pneumonia and therefore, diminishing the high mortality that is still associated with it. The search strategy for the review was broad to locate all studies in which some type of molecule was used as an alternative treatment or blockade of the toxin. The articles reviewed focused on experimental and laboratory studies and provide reduced information about the behavior with other doses or routes of inoculation. Most studies do not provide information on survival rates and limit themselves to studying the characteristics of the organs or tissues involved. One of the limitations of this systematic review is the lack of homogeneity of the studies. We did not find comparable studies in molecules, doses, inoculation routes, etc. that allow a statistical comparison of the results or meta-analysis. However, most of the studies reviewed has a good level of confidence. The clinical efficacy of the molecules in the treatment of pneumococcal infections will depend on several factors such as the pharmacokinetic/pharmacodynamic parameters of the molecule used for treating the infection, and the infection site (i.e. Central Nervous System is a difficult place for the molecules to penetrate). Indeed, some compounds have a good perspective for inhibition of pore formation and apoptosis, such as polyphenols and sterols, which also exhibit anti-inflammatory properties. With respect to hyperpermeability, it appears that it can be reduced by thioethers and some antimicrobial peptides. However, although some of these compounds have a great potential to be implemented as adjuvants in the treatment of pneumococcal infections, others (like peptides revised in this study), require future clinical trials using different combinations and deeper investigation. Hopefully, recent research related to the SARS-CoV-2 pandemic will help to understand the effect of these molecules with respect to ARDS in pneumococcal pneumonia.

Other limitation of this review was that surprisingly, for some molecules, (like cations and cholesterol), of which we did not find PLY-related studies in animal models, there are a considerable number of clinical trials studies related with pneumonia. We considered that this need exhaustive testing and are out of score of this review.

In summary, although the clinical relevance of this review is limited due to the lack of clinical tests in humans, some molecules have already been clinically tested and therefore could be safe for use in pneumococcal infections.

Supporting information

S1 Fig. Pneumolysin (PLY) action mechanisms in the different body compartments. (a)

Toxin interactions with cellular membranes (I), human immune system (II), and endothelium (III) are depicted. (b) The pneumococcus spreading routes and the main protein domains relevant to the structure of PLY are indicated.

(TIF)

S1 Table. PRISMA 2020 checklist. (DOCX)

S2 Table. Data extraction items. (DOCX)

S3 Table. Study quality assessment based on the OHAT risk assessment. (DOCX)

S4 Table. Articles and number of clinical trials. (DOCX)

S5 Table. Chemical structures of molecules with effects against PLY. (DOCX)

Acknowledgments

We acknowledge Dr. Pilar García (IPLA, CSIC, Spain) for critical review of manuscript.

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References

- Feldman C.; Anderson R. The Role of Streptococcus Pneumoniae in Community-Acquired Pneumonia. Semin Respir Crit Care Med 2020, 41, 455–469, https://doi.org/10.1055/s-0040-1702193 PMID: 32629486
- Mitchell A.M.; Mitchell T.J. Streptococcus Pneumoniae: Virulence Factors and Variation. *Clin Microbiol Infect* 2010, 16, 411–418, https://doi.org/10.1111/j.1469-0691.2010.03183.x PMID: 20132250
- 3. Briles D.E.; Paton J.C.; Mukerji R.; Swiatlo E.; Crain M.J. Pneumococcal Vaccines. *Microbiol Spectr* 2019, 7, https://doi.org/10.1128/microbiolspec.GPP3-0028-2018 PMID: 31858954

- Kadioglu A.; Weiser J.N.; Paton J.C.; Andrew P.W. The Role of Streptococcus Pneumoniae Virulence Factors in Host Respiratory Colonization and Disease. *Nat Rev Microbiol* 2008, 6, 288–301, <u>https://doi.org/10.1038/nrmicro1871</u> PMID: <u>18340341</u>
- Rowe H.M.; Karlsson E.; Echlin H.; Chang T.-C.; Wang L.; van Opijnen T.; et al. Bacterial Factors Required for Transmission of Streptococcus Pneumoniae in Mammalian Hosts. *Cell Host Microbe* 2019, 25, 884–891.e6, https://doi.org/10.1016/j.chom.2019.04.012 PMID: 31126758
- Yu Y.; Chang D.; Xu H.; Zhang X.; Pan L.; Xu C.; et al. The Virulence of Streptococcus Pneumoniae Partially Depends on DprA. *Braz J Microbiol* 2017, 48, 225–231, https://doi.org/10.1016/j.bjm.2016. 10.019 PMID: 28011228
- Subramanian K.; Iovino F.; Tsikourkitoudi V.; Merkl P.; Ahmed S.; Berry S.B.; et al. Mannose Receptor-Derived Peptides Neutralize Pore-Forming Toxins and Reduce Inflammation and Development of Pneumococcal Disease. *EMBO Mol Med* 2020, 12, e12695, <u>https://doi.org/10.15252/emmm.</u> 202012695 PMID: 32985105
- Subramanian K.; Neill D.R.; Malak H.A.; Spelmink L.; Khandaker S.; Dalla Libera Marchiori G.; et al. Pneumolysin Binds to the Mannose Receptor C Type 1 (MRC-1) Leading to Anti-Inflammatory Responses and Enhanced Pneumococcal Survival. *Nat Microbiol* 2019, 4, 62–70, https://doi.org/10. 1038/s41564-018-0280-x PMID: 30420782
- Surve M.V.; Bhutda S.; Datey A.; Anil A.; Rawat S.; Pushpakaran A.; et al. Heterogeneity in Pneumolysin Expression Governs the Fate of Streptococcus Pneumoniae during Blood-Brain Barrier Trafficking. *PLoS Pathog* 2018, 14, e1007168, https://doi.org/10.1371/journal.ppat.1007168 PMID: 30011336
- Mellroth P.; Daniels R.; Eberhardt A.; Rönnlund D.; Blom H.; Widengren J.; et al. LytA, Major Autolysin of Streptococcus Pneumoniae, Requires Access to Nascent Peptidoglycan. *J Biol Chem* 2012, 287, 11018–11029, https://doi.org/10.1074/jbc.M111.318584 PMID: 22334685
- Flores-Kim J.; Dobihal G.S.; Bernhardt T.G.; Rudner D.Z. WhyD Tailors Surface Polymers to Prevent Premature Bacteriolysis and Direct Cell Elongation in Streptococcus Pneumoniae. *Elife* 2022, 11, e76392, https://doi.org/10.7554/eLife.76392 PMID: 35593695
- Mario C.; Sandra M.; Federico I.; Priyanka N.; Laura P.; Nyunt W.S.; et al. Immunomodulatory Effects of Pneumococcal Extracellular Vesicles on Cellular and Humoral Host Defenses. *mBio* 2018, 9, e00559–18, https://doi.org/10.1128/mBio.00559-18 PMID: 29636428
- Surve M.V.; Banerjee A. Cell-to-Cell Phenotypic Heterogeneity in Pneumococcal Pathogenesis. *Future Microbiol* 2019, 14, 647–651, https://doi.org/10.2217/fmb-2019-0096 PMID: 31148481
- Greene N.G.; Narciso A.R.; Filipe S.R.; Camilli A. Peptidoglycan Branched Stem Peptides Contribute to Streptococcus Pneumoniae Virulence by Inhibiting Pneumolysin Release. *PLoS Pathog* 2015, 11, e1004996, https://doi.org/10.1371/journal.ppat.1004996 PMID: 26114646
- Martin V.; M., B.R.; Estefania M.; Katharina van P.; Özkan Y.; Werner K.; et al. Membrane Perforation by the Pore-Forming Toxin Pneumolysin. *Proceedings of the National Academy of Sciences* 2019, 116, 13352–13357, https://doi.org/10.1073/pnas.1904304116 PMID: 31209022
- Jefferies J.M.C.; Johnston C.H.G.; Kirkham L.-A.S.; Cowan G.J.M.; Ross K.S.; Smith A.; et al. Presence of Nonhemolytic Pneumolysin in Serotypes of Streptococcus Pneumoniae Associated with Disease Outbreaks. *Journal of Infectious Diseases* 2007, 196, 936–944, https://doi.org/10.1086/520091 PMID: 17703426
- Badgujar D.C.; Anil A.; Green A.E.; Surve M.V.; Madhavan S.; Beckett A.; et al. Structural Insights into Loss of Function of a Pore Forming Toxin and Its Role in Pneumococcal Adaptation to an Intracellular Lifestyle. *PLoS Pathog* 2020, 16, e1009016–. <u>https://doi.org/10.1371/journal.ppat.1009016</u> PMID: 33216805
- Jacques L.C.; Panagiotou S.; Baltazar M.; Senghore M.; Khandaker S.; Xu R.; et al. Increased Pathogenicity of Pneumococcal Serotype 1 Is Driven by Rapid Autolysis and Release of Pneumolysin. *Nat Commun* 2020, 11, 1892, https://doi.org/10.1038/s41467-020-15751-6 PMID: 32312961
- Chaguza C.; Yang M.; Jacques L.C.; Bentley S.D.; Kadioglu A. Serotype 1 Pneumococcus: Epidemiology, Genomics, and Disease Mechanisms. *Trends Microbiol* 2022, 30, 581–592, <u>https://doi.org/10.1016/j.tim.2021.11.007</u> PMID: 34949516
- van Pee K.; Neuhaus A.; D'Imprima E.; Mills D.J.; Kühlbrandt W.; Yildiz Ö. CryoEM Structures of Membrane Pore and Prepore Complex Reveal Cytolytic Mechanism of Pneumolysin. *Elife* 2017, 6, <u>https://</u> doi.org/10.7554/eLife.23644 PMID: 28323617
- Marshall J.E.; Faraj B.H.A.; Gingras A.R.; Lonnen R.; Sheikh M.A.; El-Mezgueldi M.; et al. The Crystal Structure of Pneumolysin at 2.0 Å Resolution Reveals the Molecular Packing of the Pre-Pore Complex. *Sci Rep* 2015, 5, 13293, https://doi.org/10.1038/srep13293 PMID: 26333773
- Lawrence S.L.; Feil S.C.; Morton C.J.; Farrand A.J.; Mulhern T.D.; Gorman M.A.; et al. Crystal Structure of Streptococcus Pneumoniae Pneumolysin Provides Key Insights into Early Steps of Pore Formation. *Sci Rep* 2015, 5, 14352, https://doi.org/10.1038/srep14352 PMID: 26403197

- Rai P.; He F.; Kwang J.; Engelward B.P.; Chow V.T.K. Pneumococcal Pneumolysin Induces DNA Damage and Cell Cycle Arrest. *Sci Rep* 2016, 6, 22972, https://doi.org/10.1038/srep22972 PMID: 27026501
- 24. Karmakar M.; Katsnelson M.; Malak H.A.; Greene N.G.; Howell S.J.; Hise A.G.; et al. Neutrophil IL-1β Processing Induced by Pneumolysin Is Mediated by the NLRP3/ASC Inflammasome and Caspase-1 Activation and Is Dependent on K+ Efflux. *J Immunol* 2015, 194, 1763–1775, <u>https://doi.org/10.4049/</u> jimmunol.1401624 PMID: 25609842
- González-Juarbe N.; Gilley R.P.; Hinojosa C.A.; Bradley K.M.; Kamei A.; Gao G.; et al. Pore-Forming Toxins Induce Macrophage Necroptosis during Acute Bacterial Pneumonia. *PLoS Pathog* 2015, 11, e1005337, https://doi.org/10.1371/journal.ppat.1005337 PMID: 26659062
- Nerlich A.; Mieth M.; Letsiou E.; Fatykhova D.; Zscheppang K.; Imai-Matsushima A.; et al. Pneumolysin Induced Mitochondrial Dysfunction Leads to Release of Mitochondrial DNA. *Sci Rep* 2018, 8, 182, https://doi.org/10.1038/s41598-017-18468-7 PMID: 29317705
- Letsiou E.; Teixeira Alves L.G.; Fatykhova D.; Felten M.; Mitchell T.J.; Müller-Redetzky H.C.; et al. Microvesicles Released from Pneumolysin-Stimulated Lung Epithelial Cells Carry Mitochondrial Cargo and Suppress Neutrophil Oxidative Burst. *Sci Rep* 2021, 11, 9529, https://doi.org/10.1038/ s41598-021-88897-y PMID: 33953279
- Dogan S.; Zhang Q.; Pridmore A.C.; Mitchell T.J.; Finn A.; Murdoch C. Pneumolysin-Induced CXCL8 Production by Nasopharyngeal Epithelial Cells Is Dependent on Calcium Flux and MAPK Activation via Toll-like Receptor 4. *Microbes Infect* 2011, 13, 65–75, https://doi.org/10.1016/j.micinf.2010.10.003 PMID: 20974276
- Lemon J.K.; Weiser J.N. Degradation Products of the Extracellular Pathogen Streptococcus Pneumoniae Access the Cytosol via Its Pore-Forming Toxin. *mBio* 2015, 6, https://doi.org/10.1128/mBio. 02110-14 PMID: 25604786
- Malley R.; Henneke P.; Morse S.C.; Cieslewicz M.J.; Lipsitch M.; Thompson C.M.; et al. Recognition of Pneumolysin by Toll-like Receptor 4 Confers Resistance to Pneumococcal Infection. *Proc Natl Acad Sci U S A* 2003, 100, 1966–1971, https://doi.org/10.1073/pnas.0435928100 PMID: 12569171
- Srivastava A.; Henneke P.; Visintin A.; Morse S.C.; Martin V.; Watkins C.; et al. The Apoptotic Response to Pneumolysin Is Toll-like Receptor 4 Dependent and Protects against Pneumococcal Disease. *Infect Immun* 2005, 73, 6479–6487, https://doi.org/10.1128/IAI.73.10.6479-6487.2005 PMID: 16177320
- McNeela E.A.; Burke A.; Neill D.R.; Baxter C.; Fernandes V.E.; Ferreira D.; et al. Pneumolysin Activates the NLRP3 Inflammasome and Promotes Proinflammatory Cytokines Independently of TLR4. PLoS Pathog 2010, 6, e1001191, https://doi.org/10.1371/journal.ppat.1001191 PMID: 21085613
- Cockeran R.; Theron A.J.; Steel H.C.; Matlola N.M.; Mitchell T.J.; Feldman C.; et al. Proinflammatory Interactions of Pneumolysin with Human Neutrophils. *J Infect Dis* 2001, 183, 604–611, https://doi.org/ 10.1086/318536 PMID: 11170986
- Cole J.; Angyal A.; Emes R.D.; Mitchell T.J.; Dickman M.J.; Dockrell D.H. Pneumolysin Is Responsible for Differential Gene Expression and Modifications in the Epigenetic Landscape of Primary Monocyte Derived Macrophages. *Front Immunol* 2021, 12. <u>https://doi.org/10.3389/fimmu.2021.573266</u> PMID: 34046027
- Chaudhry H.; Zhou J.; Zhong Y.; Ali M.M.; McGuire F.; Nagarkatti P.S.; et al. Role of Cytokines as a Double-Edged Sword in Sepsis. *In Vivo* 2013, 27, 669–684. PMID: 24292568
- 36. Witzenrath M.; Gutbier B.; Hocke A.C.; Schmeck B.; Hippenstiel S.; Berger K.; et al. Role of Pneumolysin for the Development of Acute Lung Injury in Pneumococcal Pneumonia. *Crit Care Med* 2006, 34, 1947–1954, https://doi.org/10.1097/01.CCM.0000220496.48295.A9 PMID: 16715037
- **37.** Lucas R.; Yang G.; Gorshkov B.A.; Zemskov E.A.; Sridhar S.; Umapathy N.S.; et al. Protein Kinase Cα and Arginase I Mediate Pneumolysin-Induced Pulmonary Endothelial Hyperpermeability. *Am J Respir Cell Mol Biol* 2012, 47, 445–453, https://doi.org/10.1165/rcmb.2011-0332OC PMID: 22582175
- Kostadinova E.; Chaput C.; Gutbier B.; Lippmann J.; Sander L.E.; Mitchell T.J.; et al. NLRP3 Protects Alveolar Barrier Integrity by an Inflammasome-Independent Increase of Epithelial Cell Adherence. *Sci Rep* 2016, 6, 30943, https://doi.org/10.1038/srep30943 PMID: 27476670
- Lucas R.; Czikora I.; Sridhar S.; Zemskov E.; Gorshkov B.; Siddaramappa U.; et al. Mini-Review: Novel Therapeutic Strategies to Blunt Actions of Pneumolysin in the Lungs. *Toxins (Basel)* 2013, 5, 1244–1260, https://doi.org/10.3390/toxins5071244 PMID: 23860351
- Zafar M.A.; Wang Y.; Hamaguchi S.; Weiser J.N. Host-to-Host Transmission of Streptococcus Pneumoniae Is Driven by Its Inflammatory Toxin, Pneumolysin. *Cell Host Microbe* 2017, 21, 73–83, https://doi.org/10.1016/j.chom.2016.12.005 PMID: 28081446

- Weiser J.N.; Ferreira D.M.; Paton J.C. Streptococcus Pneumoniae: Transmission, Colonization and Invasion. Nat Rev Microbiol 2018, 16, 355–367, https://doi.org/10.1038/s41579-018-0001-8 PMID: 29599457
- Hirst R.A.; Kadioglu A.; O'callaghan C.; Andrew P.W. The Role of Pneumolysin in Pneumococcal Pneumonia and Meningitis. *Clin Exp Immunol* 2004, 138, 195–201, https://doi.org/10.1111/j.1365-2249.2004.02611.x PMID: 15498026
- Anderson R.; Nel J.G.; Feldman C. Multifaceted Role of Pneumolysin in the Pathogenesis of Myocardial Injury in Community-Acquired Pneumonia. *Int J Mol Sci* 2018, 19, https://doi.org/10.3390/ ijms19041147 PMID: 29641429
- Alhamdi Y.; Neill D.R.; Abrams S.T.; Malak H.A.; Yahya R.; Barrett-Jolley R.; et al. Circulating Pneumolysin Is a Potent Inducer of Cardiac Injury during Pneumococcal Infection. *PLoS Pathog* 2015, 11, e1004836, https://doi.org/10.1371/journal.ppat.1004836 PMID: 25973949
- 45. Jahn K.; Handtke S.; Palankar R.; Weißmüller S.; Nouailles G.; Kohler T.P.; et al. Pneumolysin Induces Platelet Destruction, Not Platelet Activation, Which Can Be Prevented by Immunoglobulin Preparations in Vitro. *Blood Adv* 2020, 4, 6315–6326, https://doi.org/10.1182/bloodadvances. 2020002372 PMID: 33351126
- 46. Oehmcke-Hecht S.; Maletzki C.; Surabhi S.; Siemens N.; Khaimov V.; John L.M.; et al. Procoagulant Activity of Blood and Microvesicles Is Disturbed by Pneumococcal Pneumolysin, Which Interacts with Coagulation Factors. J Innate Immun 2023, 136–152, https://doi.org/10.1159/000525479 PMID: 35843205
- 47. Matanock A.; Lee G.; Gierke R.; Kobayashi M.; Leidner A.; Pilishvili T. Use of 13-Valent Pneumococcal Conjugate Vaccine and 23-Valent Pneumococcal Polysaccharide Vaccine Among Adults Aged ≥65 Years: Updated Recommendations of the Advisory Committee on Immunization Practices. *MMWR Morb Mortal Wkly Rep* 2019, 68, 1069–1075, https://doi.org/10.15585/mmwr.mm6846a5 PMID: 31751323
- Kobayashi M.; Farrar J.L.; Gierke R.; Britton A.; Childs L.; Leidner A.J.; et al. Use of 15-Valent Pneumococcal Conjugate Vaccine and 20-Valent Pneumococcal Conjugate Vaccine Among U.S. Adults: Updated Recommendations of the Advisory Committee on Immunization Practices—United States, 2022. MMWR Morb Mortal Wkly Rep 2022, 71, 109–117, https://doi.org/10.15585/mmwr.mm7104a1 PMID: 35085226
- 49. Metlay J.P.; Waterer G.W.; Long A.C.; Anzueto A.; Brozek J.; Crothers K.; et al. Diagnosis and Treatment of Adults with Community-Acquired Pneumonia. An Official Clinical Practice Guideline of the American Thoracic Society and Infectious Diseases Society of America. Am J Respir Crit Care Med 2019, 200, e45–e67, https://doi.org/10.1164/rccm.201908-1581ST PMID: 31573350
- Li L.; Ma J.; Yu Z.; Li M.; Zhang W.; Sun H. Epidemiological Characteristics and Antibiotic Resistance Mechanisms of Streptococcus Pneumoniae: An Updated Review. *Microbiol Res* 2023, 266, 127221, https://doi.org/10.1016/j.micres.2022.127221 PMID: 36244081
- Lo S.W.; Gladstone R.A.; van Tonder A.J.; Lees J.A.; du Plessis M.; Benisty R.; et al. Pneumococcal Lineages Associated with Serotype Replacement and Antibiotic Resistance in Childhood Invasive Pneumococcal Disease in the Post-PCV13 Era: An International Whole-Genome Sequencing Study. Lancet Infect Dis 2019, 19, 759–769, https://doi.org/10.1016/S1473-3099(19)30297-X PMID: 31196809
- Daniels C.C.; Rogers P.D.; Shelton C.M. A Review of Pneumococcal Vaccines: Current Polysaccharide Vaccine Recommendations and Future Protein Antigens. *J Pediatr Pharmacol Ther* 2016, 21, 27–35, https://doi.org/10.5863/1551-6776-21.1.27 PMID: 26997927
- Darrieux M.; Goulart C.; Briles D.; Leite L.C. de C. Current Status and Perspectives on Protein-Based Pneumococcal Vaccines. *Crit Rev Microbiol* 2015, 41, 190–200, https://doi.org/10.3109/1040841X. 2013.813902 PMID: 23895377
- Moffitt K.; Malley R. Rationale and Prospects for Novel Pneumococcal Vaccines. Hum Vaccin Immunother 2016, 12, 383–392, https://doi.org/10.1080/21645515.2015.1087625 PMID: 26535755
- Morais V.; Texeira E.; Suarez N. Next-Generation Whole-Cell Pneumococcal Vaccine. Vaccines (Basel) 2019, 7, https://doi.org/10.3390/vaccines7040151 PMID: 31623286
- 56. Kim G.-L.; Seon S.-H.; Rhee D.-K. Pneumonia and Streptococcus Pneumoniae Vaccine. Arch Pharm Res 2017, 40, 885–893, https://doi.org/10.1007/s12272-017-0933-y PMID: 28735461
- 57. Nishimoto A.T.; Rosch J.W.; Tuomanen E.I. Pneumolysin: Pathogenesis and Therapeutic Target. *Front Microbiol* 2020, 11, 1543, https://doi.org/10.3389/fmicb.2020.01543 PMID: 32714314
- Li H.; Zhao X.; Deng X.; Wang J.; Song M.; Niu X.; et al. Insights into Structure and Activity of Natural Compound Inhibitors of Pneumolysin. *Sci Rep* 2017, 7, 42015, https://doi.org/10.1038/srep42015 PMID: 28165051

- Anderson R.; Feldman C. Pneumolysin as a Potential Therapeutic Target in Severe Pneumococcal Disease. J Infect 2017, 74, 527–544, https://doi.org/10.1016/j.jinf.2017.03.005 PMID: 28322888
- Page M.J.; McKenzie J.E.; Bossuyt P.M.; Boutron I.; Hoffmann T.C.; Mulrow C.D.; et al. The PRISMA 2020 Statement: An Updated Guideline for Reporting Systematic Reviews. *BMJ* 2021, 372, n71, https://doi.org/10.1136/bmj.n71 PMID: 33782057
- **61.** National Toxicology Program OHAT Risk of Bias Rating Tool for Human and Animal Studies. *OHAT Risk of Bias Tool* 2015, US Departm, 1–37.
- Pafilis E.; Bērziņš R.; Jensen L.J. EXTRACT 2.0: Text-Mining-Assisted Interactive Annotation of Biomedical Named Entities and Ontology Terms. *bioRxiv* 2017, 111088, https://doi.org/10.1101/111088
- Shannon P.; Markiel A.; Ozier O.; Baliga N.S.; Wang J.T.; Ramage D. Cytoscape: A Software Environment for Integrated Models of Biomolecular Interaction Networks. *Genome Res* 2003, 13, 2498–2504, https://doi.org/10.1101/gr.1239303 PMID: 14597658
- Le C.-F.; Yusof M.Y.M.; Hassan M.A.A.; Lee V.S.; Isa D.M.; Sekaran S.D. In Vivo Efficacy and Molecular Docking of Designed Peptide That Exhibits Potent Antipneumococcal Activity and Synergises in Combination with Penicillin. *Sci Rep* 2015, 5, 11886, <u>https://doi.org/10.1038/srep11886</u> PMID: 26156658
- Subramaniam Y.; Subbiah R.; Balan L.; Subban K. Bioprospecting of Bioactive Metabolites from Monochaetia Karstenii. *J Pure Appl Microbiol* 2020, 14, 1557–1566, <u>https://doi.org/10.22207/JPAM.</u> 14.2.54
- 66. Hupp S.; Ribes S.; Seele J.; Bischoff C.; Förtsch C.; Maier E.; et al. Magnesium Therapy Improves Outcome in Streptococcus Pneumoniae Meningitis by Altering Pneumolysin Pore Formation. Br J Pharmacol 2017, 174, 4295–4307, https://doi.org/10.1111/bph.14027 PMID: 28888095
- Marquart M.E.; Monds K.S.; McCormick C.C.; Dixon S.N.; Sanders M.E.; Reed J.M.; et al. Cholesterol as Treatment for Pneumococcal Keratitis: Cholesterol-Specific Inhibition of Pneumolysin in the Cornea. *Invest Ophthalmol Vis Sci* 2007, 48, 2661–2666, <u>https://doi.org/10.1167/iovs.07-0017</u> PMID: 17525197
- Green S.N.; Sanders M.; Moore Q.C. 3rd; Norcross E.W.; Monds K.S.; Caballero A.R.; et al. Protection from Streptococcus Pneumoniae Keratitis by Passive Immunization with Pneumolysin Antiserum. *Invest Ophthalmol Vis Sci* 2008, 49, 290–294, https://doi.org/10.1167/iovs.07-0492 PMID: 18172105
- Chang S.-F.; Chen C.-N.; Lin J.-C.; Wang H.-E.; Mori S.; Li J.-J.; et al. Truncated Pneumolysin from Streptococcus Pneumoniae as a TLR4-Antagonizing New Drug for Chronic Inflammatory Conditions. *Cells* 2020, 9, https://doi.org/10.3390/cells9051183 PMID: 32397494
- 70. Kaur R.; Surendran N.; Ochs M.; Pichichero M.E. Human Antibodies to PhtD, PcpA, and Ply Reduce Adherence to Human Lung Epithelial Cells and Murine Nasopharyngeal Colonization by Streptococcus Pneumoniae. *Infect Immun* 2014, 82, 5069–5075, <u>https://doi.org/10.1128/IAI.02124-14</u> PMID: 25245804
- Otieno W.; Liu C.; Ji Y. Aloe-Emodin-Mediated Photodynamic Therapy Attenuates Sepsis-Associated Toxins in Selected Gram-Positive Bacteria In Vitro. *J Microbiol Biotechnol* 2021, 31, 1200–1209, https://doi.org/10.4014/jmb.2105.05024 PMID: 34319262
- Barbieri R.; Coppo E.; Marchese A.; Daglia M.; Sobarzo-Sánchez E.; Nabavi S.F.; et al. Phytochemicals for Human Disease: An Update on Plant-Derived Compounds Antibacterial Activity. *Microbiol Res* 2017, 196, 44–68, https://doi.org/10.1016/j.micres.2016.12.003 PMID: 28164790
- 73. He Y.-Q.; Zhou C.-C.; Yu L.-Y.; Wang L.; Deng J.-L.; Tao Y.-L.; et al. Natural Product Derived Phytochemicals in Managing Acute Lung Injury by Multiple Mechanisms. *Pharmacol Res* 2021, 163, 105224, https://doi.org/10.1016/j.phrs.2020.105224 PMID: 33007416
- 74. Bernini R.; Velotti F. Natural Polyphenols as Immunomodulators to Rescue Immune Response Homeostasis: Quercetin as a Research Model against Severe COVID-19. *Molecules* 2021, 26.
- 75. Zhao X.; Li H.; Wang J.; Guo Y.; Liu B.; Deng X.; et al. Verbascoside Alleviates Pneumococcal Pneumonia by Reducing Pneumolysin Oligomers. *Mol Pharmacol* 2016, 89, 376–387, <u>https://doi.org/10.1124/mol.115.100610 PMID: 26700563</u>
- 76. Song M.; Li L.; Li M.; Cha Y.; Deng X.; Wang J. Apigenin Protects Mice from Pneumococcal Pneumonia by Inhibiting the Cytolytic Activity of Pneumolysin. *Fitoterapia* 2016, 115, 31–36, <u>https://doi.org/10.1016/j.fitote.2016.09.017</u> PMID: 27693741
- Li S.; Lv Q.; Sun X.; Tang T.; Deng X.; Yin Y.; et al. Acacetin Inhibits Streptococcus Pneumoniae Virulence by Targeting Pneumolysin. *J Pharm Pharmacol* 2020, 72, 1092–1100, https://doi.org/10.1111/jphp.13279 PMID: 32390150
- Zhao X.; Liu B.; Liu S.; Wang L.; Wang J. Anticytotoxin Effects of Amentoflavone to Pneumolysin. Biol Pharm Bull 2017, 40, 61–67, https://doi.org/10.1248/bpb.b16-00598 PMID: 28049950

- 79. Zhao X.; Zhou Y.; Wang G.; Shi D.; Zha Y.; Yi P.; et al. Morin Moderates the Biotoxicity of Pneumococcal Pneumolysin by Weakening the Oligomers' Formation. *Chem Pharm Bull (Tokyo)* 2017, 65, 538– 544, https://doi.org/10.1248/cpb.c16-00999 PMID: 28566646
- Song M.; Teng Z.; Li M.; Niu X.; Wang J.; Deng X. Epigallocatechin Gallate Inhibits Streptococcus Pneumoniae Virulence by Simultaneously Targeting Pneumolysin and Sortase A. J Cell Mol Med 2017, 21, 2586–2598, https://doi.org/10.1111/jcmm.13179 PMID: 28402019
- Lv Q.; Zhang P.; Quan P.; Cui M.; Liu T.; Yin Y.; et al. Quercetin, a Pneumolysin Inhibitor, Protects Mice against Streptococcus Pneumoniae Infection. *Microb Pathog* 2020, 140, 103934, <u>https://doi.org/10.1016/j.micpath.2019.103934</u> PMID: 31862394
- Lv H.; Fang T.; Kong F.; Wang J.; Deng X.; Yu Q.; et al. Dryocrassin ABBA Ameliorates Streptococcus Pneumoniae-Induced Infection in Vitro through Inhibiting Streptococcus Pneumoniae Growth and Neutralizing Pneumolysin Activity. *Microb Pathog* 2021, 150, 104683, https://doi.org/10.1016/j. micpath.2020.104683 PMID: 33309685
- Maatsola S.; Kurkinen S.; Engström M.T.; Nyholm T.K.M.; Pentikäinen O.; Salminen J.-P.; et al. Inhibition of Pneumolysin Cytotoxicity by Hydrolysable Tannins. *Antibiotics (Basel)* 2020, 9, 930, <u>https://doi.org/10.3390/antibiotics9120930</u> PMID: 33371182
- Zhao X.; Zhou Y.; Wang L.; Li M.; Shi D.; Li D.; Wang J. Shikonin Alleviates the Biotoxicity Produced by Pneumococcal Pneumolysin. *Life Sci* 2017, 177, 1–7, <u>https://doi.org/10.1016/j.lfs.2017.04.002</u> PMID: 28385613
- Song M.; Lu G.; Li M.; Deng X.; Wang J. Juglone Alleviates Pneumolysin-Induced Human Alveolar Epithelial Cell Injury via Inhibiting the Hemolytic Activity of Pneumolysin. *Antonie Van Leeuwenhoek* 2017, 110, 1069–1075, https://doi.org/10.1007/s10482-017-0880-0 PMID: 28451868
- Xu Y.; Wei L.; Wang Y.; Ding L.; Guo Y.; Sun X.; et al. Inhibitory Effect of the Traditional Chinese Medicine Ephedra Sinica Granules on Streptococcus Pneumoniae Pneumolysin. *Biol Pharm Bull* 2020, 43, 994–999, https://doi.org/10.1248/bpb.b20-00034 PMID: 32475921
- Guo T.; Guo Y.; Liu Q.; Xu Y.; Wei L.; Wang Z.; et al. The TCM Prescription Ma-Xing-Shi-Gan-Tang Inhibits Streptococcus Pneumoniae Pathogenesis by Targeting Pneumolysin. *J Ethnopharmacol* 2021, 275, 114133, https://doi.org/10.1016/j.jep.2021.114133 PMID: 33892068
- He W.; Ma J.; Chen Y.; Jiang X.; Wang Y.; Shi T.; et al. Ephedrine Hydrochloride Protects Mice from Staphylococcus Aureus-Induced Peritonitis. *Am J Transl Res* 2018, 10, 670–683. PMID: 29636858
- 89. Zhou Y.; Guo Y.; Sun X.; Ding R.; Wang Y.; Niu X.; et al. Application of Oleanolic Acid and Its Analogues in Combating Pathogenic Bacteria In Vitro/Vivo by a Two-Pronged Strategy of β-Lactamases and Hemolysins. ACS Omega 2020, 5, 11424–11438, https://doi.org/10.1021/acsomega.0c00460 PMID: 32478231
- 90. Zhang Z.Q.A.-Z.Q.A.-Y.G.A.-H.Z.A.-Q.Y.A.-P. Betulin Attenuates Pneumolysin-induced Cell Injury and DNA Damage. J Appl Microbiol 2021, v. 130, 843–851–2021 v.130 no.3, <u>https://doi.org/10.1111/jam.14769 PMID: 32621771</u>
- Ding R.; Zhang Y.; Xu X.; Hou Y.; Nie J.; Deng X.; et al. Inhibitory Effect of Hederagenin on Streptococcus Pneumoniae Pneumolysin in Vitro. *Microbes Infect* 2022, 24, 104888, <u>https://doi.org/10.1016/j.</u> micinf.2021.104888 PMID: 34547436
- 92. di Pierro F.; Derosa G.; Maffioli P.; Bertuccioli A.; Togni S.; Riva A.; et al. Possible Therapeutic Effects of Adjuvant Quercetin Supplementation Against Early-Stage COVID-19 Infection: A Prospective, Randomized, Controlled, and Open-Label Study. Int J Gen Med 2021, 14, 2359–2366. <u>https://doi.org/10. 2147/IJGM.S318720 PMID: 34135619</u>
- 93. Li H.; Zhao X.; Wang J.; Dong Y.; Meng S.; Li R.; et al. β-Sitosterol Interacts with Pneumolysin to Prevent Streptococcus Pneumoniae Infection. *Sci Rep* 2015, 5, 17668, <u>https://doi.org/10.1038/srep17668</u> PMID: 26631364
- 94. Fraile L.; Crisci E.; Córdoba L.; Navarro M.A.; Osada J.; Montoya M. Immunomodulatory Properties of Beta-Sitosterol in Pig Immune Responses. Int Immunopharmacol 2012, 13, 316–321, https://doi.org/ 10.1016/j.intimp.2012.04.017 PMID: 22595193
- **95.** Kasirzadeh S.; Ghahremani M.H.; Setayesh N.; Jeivad F.; Shadboorestan A.; Taheri A.; et al. β-Sitosterol Alters the Inflammatory Response in CLP Rat Model of Sepsis by Modulation of NF*κ*B Signaling. *Biomed Res Int* 2021, 2021, 5535562, https://doi.org/10.1155/2021/5535562 PMID: 33997001
- 96. Statt S.; Ruan J.-W.; Huang C.-T.; Wu R.; Kao C.-Y. Lipidome and Transcriptome Profiling of Pneumolysin Intoxication Identifies Networks Involved in Statin-Conferred Protection of Airway Epithelial Cells. *Sci Rep* 2015, 5, 10624, https://doi.org/10.1038/srep10624 PMID: 26023727
- 97. Statt S.; Ruan J.-W.; Hung L.-Y.; Chang C.-Y.; Huang C.-T.; Lim J.H.; et al. Statin-Conferred Enhanced Cellular Resistance against Bacterial Pore-Forming Toxins in Airway Epithelial Cells. Am J Respir Cell Mol Biol 2015, 53, 689–702, https://doi.org/10.1165/rcmb.2014-0391OC PMID: 25874372

- Boyd A.R.; Hinojosa C.A.; Rodriguez P.J.; Orihuela C.J. Impact of Oral Simvastatin Therapy on Acute Lung Injury in Mice during Pneumococcal Pneumonia. *BMC Microbiol* 2012, 12, 73, https://doi.org/10. 1186/1471-2180-12-73 PMID: 22587610
- 99. Rosch J.W.; Boyd A.R.; Hinojosa E.; Pestina T.; Hu Y.; Persons D.A.; et al. Statins Protect against Fulminant Pneumococcal Infection and Cytolysin Toxicity in a Mouse Model of Sickle Cell Disease. J Clin Invest 2010, 120, 627–635, https://doi.org/10.1172/JCI39843 PMID: 20093777
- 100. Guerra-De-Blas P.D.C.; Torres-González P.; Bobadilla-Del-Valle M.; Sada-Ovalle I.; Ponce-De-León-Garduño A.; Sifuentes-Osornio J. Potential Effect of Statins on Mycobacterium Tuberculosis Infection. *J Immunol Res* 2018, 2018, 7617023, https://doi.org/10.1155/2018/7617023 PMID: 30581876
- Mosepele M.; Molefe-Baikai O.J.; Grinspoon S.K.; Triant V.A. Benefits and Risks of Statin Therapy in the HIV-Infected Population. *Curr Infect Dis Rep* 2018, 20, 20, <u>https://doi.org/10.1007/s11908-018-0628-7</u> PMID: 29804227
- 102. Orlowski S.; Mourad J.-J.; Gallo A.; Bruckert E. Coronaviruses, Cholesterol and Statins: Involvement and Application for Covid-19. *Biochimie* 2021, 189, 51–64, <u>https://doi.org/10.1016/j.biochi.2021.06.</u> 005 PMID: 34153377
- 103. Majumdar S.R.; McAlister F.A.; Eurich D.T.; Padwal R.S.; Marrie T.J. Statins and Outcomes in Patients Admitted to Hospital with Community Acquired Pneumonia: Population Based Prospective Cohort Study. *BMJ* 2006, 333, 999, https://doi.org/10.1136/bmj.38992.565972.7C PMID: 17060337
- 104. Chalmers J.D.; Singanayagam A.; Murray M.P.; Hill A.T. Prior Statin Use Is Associated with Improved Outcomes in Community-Acquired Pneumonia. Am J Med 2008, 121, 1002–1007.e1, https://doi.org/ 10.1016/j.amjmed.2008.06.030 PMID: 18954848
- 105. Mortensen E.M.; Pugh M.J.; Copeland L.A.; Restrepo M.I.; Cornell J.E.; Anzueto A.; et al. Impact of Statins and Angiotensin-Converting Enzyme Inhibitors on Mortality of Subjects Hospitalised with Pneumonia. *Eur Respir J* 2008, 31, 611–617, https://doi.org/10.1183/09031936.00162006 PMID: 17959631
- 106. Thomsen R.W.; Riis A.; Kornum J.B.; Christensen S.; Johnsen S.P.; Sørensen H.T. Preadmission Use of Statins and Outcomes after Hospitalization with Pneumonia: Population-Based Cohort Study of 29,900 Patients. Arch Intern Med 2008, 168, 2081–2087, https://doi.org/10.1001/archinte.168.19. 2081 PMID: 18955636
- 107. Viasus D.; Garcia-Vidal C.; Simonetti A.F.; Dorca J.; Llopis F.; Mestre M.; et al. The Effect of Simvastatin on Inflammatory Cytokines in Community-Acquired Pneumonia: A Randomised, Double-Blind, Placebo-Controlled Trial. *BMJ Open* 2015, 5, e006251, <u>https://doi.org/10.1136/bmjopen-2014-</u>006251 PMID: 25564143
- 108. Sapey E.; Patel J.M.; Greenwood H.; Walton G.M.; Grudzinska F.; Parekh D.; et al. Simvastatin Improves Neutrophil Function and Clinical Outcomes in Pneumonia. A Pilot Randomized Controlled Clinical Trial. Am J Respir Crit Care Med 2019, 200, 1282–1293, https://doi.org/10.1164/rccm. 201812-2328OC PMID: 31206313
- 109. Fickl H.; Cockeran R.; Steel H.C.; Feldman C.; Cowan G.; Mitchell T.J.; et al. Pneumolysin-Mediated Activation of NFkappaB in Human Neutrophils Is Antagonized by Docosahexaenoic Acid. *Clin Exp Immunol* 2005, 140, 274–281, https://doi.org/10.1111/j.1365-2249.2005.02757.x PMID: 15807851
- Sandhaus S.; Swick A.G. Specialized Proresolving Mediators in Infection and Lung Injury. *Biofactors* 2021, 47, 6–18, https://doi.org/10.1002/biof.1691 PMID: 33249673
- 111. Perez-Hernandez J.; Chiurchiù V.; Perruche S.; You S. Regulation of T-Cell Immune Responses by Pro-Resolving Lipid Mediators. *Frontiers in Immunology* 2021, 12. https://doi.org/10.3389/fimmu. 2021.768133 PMID: 34868025
- 112. Basil M.C.; Levy B.D. Specialized Pro-Resolving Mediators: Endogenous Regulators of Infection and Inflammation. *Nat Rev Immunol* 2016, 16, 51–67, https://doi.org/10.1038/nri.2015.4 PMID: 26688348
- 113. Stapleton R.D.; Martin T.R.; Weiss N.S.; Crowley J.J.; Gundel S.J.; Nathens A.B.; et al. A Phase II Randomized Placebo-Controlled Trial of Omega-3 Fatty Acids for the Treatment of Acute Lung Injury. *Crit Care Med* 2011, 39, 1655–1662, https://doi.org/10.1097/CCM.0b013e318218669d PMID: 21423000
- 114. Gold D.R.; Litonjua A.A.; Carey V.J.; Manson J.E.; Buring J.E.; Lee I.-M.; et al. Lung VITAL: Rationale, Design, and Baseline Characteristics of an Ancillary Study Evaluating the Effects of Vitamin D and/or Marine Omega-3 Fatty Acid Supplements on Acute Exacerbations of Chronic Respiratory Disease, Asthma Control, Pneumonia and Lung f. *Contemp Clin Trials* 2016, 47, 185–195, https://doi.org/10. 1016/j.cct.2016.01.003 PMID: 26784651
- 115. Sedighiyan M.; Abdollahi H.; Karimi E.; Badeli M.; Erfanian R.; Raeesi S.; et al. Omega-3 Polyunsaturated Fatty Acids Supplementation Improve Clinical Symptoms in Patients with Covid-19: A Randomised Clinical Trial. Int J Clin Pract 2021, 75, e14854, <u>https://doi.org/10.1111/ijcp.14854</u> PMID: 34516692

- 116. Witzenrath M.; Gutbier B.; Schmeck B.; Tenor H.; Seybold J.; Kuelzer R.; et al. Phosphodiesterase 2 Inhibition Diminished Acute Lung Injury in Murine Pneumococcal Pneumonia. *Crit Care Med* 2009, 37, 584–590, https://doi.org/10.1097/CCM.0b013e3181959814 PMID: 19114892
- 117. Mokra D.; Mokry J. Phosphodiesterase Inhibitors in Acute Lung Injury: What Are the Perspectives? Int J Mol Sci 2021, 22, https://doi.org/10.3390/ijms22041929 PMID: 33669167
- 118. Shigematsu M.; Koga T.; Ishimori A.; Saeki K.; Ishii Y.; Taketomi Y.; et al. Leukotriene B4 Receptor Type 2 Protects against Pneumolysin-Dependent Acute Lung Injury. *Sci Rep* 2016, 6, 34560, <u>https://doi.org/10.1038/srep34560</u> PMID: 27703200
- Kerget B.; Kerget F.; Aydın M.; Karaşahin Ö. Effect of Montelukast Therapy on Clinical Course, Pulmonary Function, and Mortality in Patients with COVID-19. J Med Virol 2022, 94, 1950–1958, <u>https://doi.org/10.1002/jmv.27552</u> PMID: 34958142
- 120. Wu H.; Ding X.; Zhao D.; Liang Y.; Ji W. Effect of Montelukast Combined with Methylprednisolone for the Treatment of Mycoplasma Pneumonia. *J Int Med Res* 2019, 47, 2555–2561, <u>https://doi.org/10. 1177/0300060518820412 PMID: 31072180</u>
- 121. Domon H.; Isono T.; Hiyoshi T.; Tamura H.; Sasagawa K.; Maekawa T.; et al. Clarithromycin Inhibits Pneumolysin Production via Downregulation of Ply Gene Transcription despite Autolysis Activation. *Microbiol Spectr* 2021, 9, e0031821, https://doi.org/10.1128/Spectrum.00318-21 PMID: 34468195
- 122. Majhi A.; Adhikary R.; Bhattacharyya A.; Mahanti S.; Bishayi B. Levofloxacin-Ceftriaxone Combination Attenuates Lung Inflammation in a Mouse Model of Bacteremic Pneumonia Caused by Multidrug-Resistant Streptococcus Pneumoniae via Inhibition of Cytolytic Activities of Pneumolysin and Autolysin. Antimicrob Agents Chemother 2014, 58, 5164–5180, https://doi.org/10.1128/AAC.03245-14 PMID: 24957840
- 123. Zarogoulidis P.; Papanas N.; Kioumis I.; Chatzaki E.; Maltezos E.; Zarogoulidis K. Macrolides: From in Vitro Anti-Inflammatory and Immunomodulatory Properties to Clinical Practice in Respiratory Diseases. *Eur J Clin Pharmacol* 2012, 68, 479–503, https://doi.org/10.1007/s00228-011-1161-x PMID: 22105373
- 124. Zusso M.; Lunardi V.; Franceschini D.; Pagetta A.; Lo R.; Stifani S.; et al. Ciprofloxacin and Levofloxacin Attenuate Microglia Inflammatory Response via TLR4/NF-KB Pathway. *J Neuroinflammation* 2019, 16, 148, https://doi.org/10.1186/s12974-019-1538-9 PMID: 31319868
- 125. Rashad A.; Nafady A.; Hassan M.H.; Mansour H.; Taya U.; Bazeed S.E.S.; et al. Therapeutic Efficacy of Macrolides in Management of Patients with Mild COVID-19. *Sci Rep* 2021, 11, 16361, <u>https://doi.org/10.1038/s41598-021-95900-z</u> PMID: 34381155
- 126. Lindenberg M.; Almeida L.; Dhillon-LaBrooy A.; Siegel E.; Henriques-Normark B.; Sparwasser T. Clarithromycin Impairs Tissue-Resident Memory and Th17 Responses to Macrolide-Resistant Streptococcus Pneumoniae Infections. J Mol Med (Berl) 2021, 99, 817–829, https://doi.org/10.1007/s00109-021-02039-5 PMID: 33595670
- 127. Mookherjee N.; Anderson M.A.; Haagsman H.P.; Davidson D.J. Antimicrobial Host Defence Peptides: Functions and Clinical Potential. *Nat Rev Drug Discov* 2020, 19, 311–332, <u>https://doi.org/10.1038/s41573-019-0058-8 PMID: 32107480</u>
- 128. Palmer N.; Maasch J.R.M.A.; Torres M.D.T.; de la Fuente-Nunez C. Molecular Dynamics for Antimicrobial Peptide Discovery. *Infect Immun* 2021, 89, <u>https://doi.org/10.1128/IAI.00703-20</u> PMID: 33558318
- 129. Lucas R.; Sridhar S.; Rick F.G.; Gorshkov B.; Umapathy N.S.; Yang G.; et al. Agonist of Growth Hormone-Releasing Hormone Reduces Pneumolysin-Induced Pulmonary Permeability Edema. *Proc Natl Acad Sci U S A* 2012, 109, 2084–2089, https://doi.org/10.1073/pnas.1121075109 PMID: 22308467
- Gutbier B.; Jiang X.; Dietert K.; Ehrler C.; Lienau J.; van Slyke P.; et al. Vasculotide Reduces Pulmonary Hyperpermeability in Experimental Pneumococcal Pneumonia. *Crit Care* 2017, 21, 274, https:// doi.org/10.1186/s13054-017-1851-6 PMID: 29132435
- 131. U.S. National Library of Medicine AV-001 for Hospitalized Patients With COVID-19 Disease (NCT05123755). *Clinical Trials.gov* 2021, November.
- 132. Schmid B.; Kredel M.; Ullrich R.; Krenn K.; Lucas R.; Markstaller K.; et al. Safety and Preliminary Efficacy of Sequential Multiple Ascending Doses of Solnatide to Treat Pulmonary Permeability Edema in Patients with Moderate-to-Severe ARDS-a Randomized, Placebo-Controlled, Double-Blind Trial. *Trials* 2021, 22, 643, https://doi.org/10.1186/s13063-021-05588-9 PMID: 34544463
- 133. N'Guessan P.D.; Schmeck B.; Ayim A.; Hocke A.C.; Brell B.; Hammerschmidt S.; et al. Streptococcus Pneumoniae R6x Induced P38 MAPK and JNK-Mediated Caspase-Dependent Apoptosis in Human Endothelial Cells. *Thromb Haemost* 2005, 94, 295–303, <u>https://doi.org/10.1160/TH04-12-0822</u> PMID: 16113818
- 134. Wolfmeier H.; Schoenauer R.; Atanassoff A.P.; Neill D.R.; Kadioglu A.; Draeger A.; et al. Ca²⁺-Dependent Repair of Pneumolysin Pores: A New Paradigm for Host Cellular Defense against Bacterial Pore-

Forming Toxins. *Biochim Biophys Acta* 2015, 1853, 2045–2054, https://doi.org/10.1016/j.bbamcr. 2014.09.005 PMID: 25219550

- 135. Franco-Vidal V.; Beurg M.; Darrouzet V.; Bébéar J.-P.; Skinner L.J.; Dulon D. Zinc Protection against Pneumolysin Toxicity on Rat Cochlear Hair Cells. *Audiol Neurootol* 2008, 13, 65–70, <u>https://doi.org/10.1159/000108763</u> PMID: 17890859
- 136. Rerksuppaphol S.; Rerksuppaphol L. A Randomized Controlled Trial of Zinc Supplementation in the Treatment of Acute Respiratory Tract Infection in Thai Children. *Pediatr Rep* 2019, 11, 7954, <u>https://doi.org/10.4081/pr.2019.7954</u> PMID: 31214301
- 137. Tan C.W.; Ho L.P.; Kalimuddin S.; Cherng B.P.Z.; Teh Y.E.; Thien S.Y.; et al. Cohort Study to Evaluate the Effect of Vitamin D, Magnesium, and Vitamin B(12) in Combination on Progression to Severe Outcomes in Older Patients with Coronavirus (COVID-19). *Nutrition* 2020, 79–80, 111017, https://doi.org/10.1016/j.nut.2020.111017 PMID: 33039952
- 138. Heller R.A.; Sun Q.; Hackler J.; Seelig J.; Seibert L.; Cherkezov A.; et al. Prediction of Survival Odds in COVID-19 by Zinc, Age and Selenoprotein P as Composite Biomarker. *Redox Biol* 2021, 38, 101764, https://doi.org/10.1016/j.redox.2020.101764 PMID: 33126054
- 139. Wippel C.; Förtsch C.; Hupp S.; Maier E.; Benz R.; Ma J.; et al. Extracellular Calcium Reduction Strongly Increases the Lytic Capacity of Pneumolysin From Streptococcus Pneumoniae in Brain Tissue. J Infect Dis 2011, 204, 930–936, https://doi.org/10.1093/infdis/jir434 PMID: 21849290
- Nerlich A.; von Wunsch Teruel I.; Mieth M.; Hönzke K.; Rückert J.C.; Mitchell T.J.; et al. Reversion of Pneumolysin-Induced Executioner Caspase Activation Redirects Cells to Survival. *J Infect Dis* 2021, 223, 1973–1983, https://doi.org/10.1093/infdis/jiaa639 PMID: 33045080
- 141. García-Suárez M. del M.; Cima-Cabal M.D.; Flórez N.; García P.; Cernuda-Cernuda R.; Astudillo A.; et al. Protection against Pneumococcal Pneumonia in Mice by Monoclonal Antibodies to Pneumolysin. *Infect Immun* 2004, 72, 4534–4540, <u>https://doi.org/10.1128/IAI.72.8.4534-4540.2004</u> PMID: 15271913
- 142. Singh S.; Kumar N.K.; Dwiwedi P.; Charan J.; Kaur R.; Sidhu P.; et al. Monoclonal Antibodies: A Review. *Curr Clin Pharmacol* 2018, 13, 85–99, https://doi.org/10.2174/ 1574884712666170809124728 PMID: 28799485
- 143. Wiebe F.; Handtke S.; Wesche J.; Schnarre A.; Palankar R.; Wolff M.; et al. Polyvalent Immunoglobulin Preparations Inhibit Pneumolysin-Induced Platelet Destruction. *Thromb Haemost* 2022, 122, 1147–1158, https://doi.org/10.1055/a-1723-1880 PMID: 34918314
- 144. Sakari M.; Laisi A.; Pulliainen A.T. Exotoxin-Targeted Drug Modalities as Antibiotic Alternatives. ACS Infect Dis 2022, 8, 433–456, https://doi.org/10.1021/acsinfecdis.1c00296 PMID: 35099182
- 145. Laterre P.-F.; Colin G.; Dequin P.-F.; Dugernier T.; Boulain T.; Azeredo da Silveira S.; et al. CAL02, a Novel Antitoxin Liposomal Agent, in Severe Pneumococcal Pneumonia: A First-in-Human, Double-Blind, Placebo-Controlled, Randomised Trial. *Lancet Infect Dis* 2019, 19, 620–630, https://doi.org/10. 1016/S1473-3099(18)30805-3 PMID: 31056427
- Somolinos F.J.; León C.; Guerrero-Aspizua S. Drug Repurposing Using Biological Networks. Processes 2021, 9.
- 147. Zhou Y.; Hou Y.; Shen J.; Huang Y.; Martin W.; Cheng F. Network-Based Drug Repurposing for Novel Coronavirus 2019-NCoV/SARS-CoV-2. *Cell Discov* 2020, 6, 14, https://doi.org/10.1038/s41421-020-0153-3 PMID: 32194980
- 148. Grau I.; Ardanuy C.; Cubero M.; Benitez M.A.; Liñares J.; Pallares R. Declining Mortality from Adult Pneumococcal Infections Linked to Children's Vaccination. J Infect 2016, 72, 439–449, https://doi. org/10.1016/j.jinf.2016.01.011 PMID: 26868606